

(12) **UK Patent Application** (19) **GB** (11) **2 332 906** (13) **A**

(43) Date of A Publication 07.07.1999

(21) Application No 9825378.4

(22) Date of Filing 19.11.1998

(30) Priority Data

(31) 06066225

(32) 20.11.1997

(33) US

(71) Applicant(s)

**F. Hoffmann-La Roche AG
(Incorporated in Switzerland)
124 Grenzacherstrasse, CH-4070 Basle, Switzerland**

(72) Inventor(s)

**Paul Shartzter Dietrich
Linda Marie Fish
Reena Khare
Douglas Kenneth Rabert
Lakshmi Sangameswaran**

(74) Agent and/or Address for Service

**Carpmaels & Ransford
43 Bloomsbury Square, LONDON, WC1A 2RA,
United Kingdom**

(51) INT CL⁶

C12N 15/12 // C07K 16/18

(52) UK CL (Edition Q)

**C3H HB7P H650 H673
U1S S2419**

(56) Documents Cited

**WO 97/01577 A WO 96/14077 A
Gene Vol. 202 1997. Chen J et. al. pages 7-14**

(58) Field of Search

**ONLINE: WPI, EPODOC, PAJ, CAS ONLINE, DGENE,
BIOSCIENCE/STN**

(54) Abstract Title

Nucleic acids encoding TTX-resistant Na channel proteins

(57) Nucleic acid sequences which encode TTX-resistant Na channel proteins derived from rat and human dorsal root ganglia are described. The products are designated PN5. The production an antiserum against a synthetic peptide from PN5 is described.

BEST AVAILABLE COPY

GB 2 332 906 A

Figure 1A: SEQ ID NO:1

```
1  GAAGTCACAG GAGTGTCTGT CAGCGAGAGG AAGAAGGGAG AGTTTACTGA
51  GTGTCTTCTG CCCCTCCTCA GGGTGAAGAT GGAGGAGAGG TACTACCCGG
101 TGATCTTCCC GGACGAGCGG AATTTCCGCC CCTTCACTTC CGACTCTCTG
151 GCTGCCATAG AGAAGCGGAT TGCTATCCAA AAGGAGAGGA AGAAGTCCAA
201 AGACAAGGCG GCAGCTGAGC CCCAGCCTCG GCCTCAGCTT GACCTAAAGG
251 CCTCCAGGAA GTTACCTAAG CTTTATGGTG ACATTCCCCC TGAGCTTGTA
301 GCGAAGCCTC TGGAAGACCT GGACCCATTC TACAAAGACC ATAAGACATT
351 CATGGTGTTG AACAAAGAAG GAACAATTTA TCGCTTCAGC GCCAAGCGGG
401 CCTTGTTTCAT TCTGGGGCCT TTTAATCCCC TCAGAAGCTT AATGATTTCG
451 ATCTCTGTCC ATTCAGTCTT TAGCATGTTC ATCATCTGCA CGGTGATCAT
501 CAACTGTATG TTCATGGCGA ATTCTATGGA GAGAAGTTTC GACAACGACA
551 TTCCCGAATA CGTCTTCATT GGGATTTATA TTTTAGAAGC TGTGATTAAA
601 ATATTGGCAA GAGGCTTCAT TGTGGATGAG TTTTCCTTCC TCCGAGATCC
651 GTGGAAGTGG CTGGACTTCA TTGTCATTGG AACAGCGATC GCAACTTGTT
701 TTCCGGGCAG CCAAGTCAAT CTTTCAGCTC TTCGTACCTT CCGAGTGTTT
751 AGAGCTCTGA AGGCGATTTT AGTTATCTCA GGTCTGAAGG TCATCGTAGG
801 TGCCCTGCTG CGCTCGGTGA AGAAGCTGGT AGACGTGATG GTCCTCACTC
851 TCTTCTGCCT CAGCATCTTT GCCCTGGTCG GTCAGCAGCT GTTCATGGGA
901 ATTCTGAACC AGAAGTGTAT TAAGCACAAC TGTGGCCCCA ACCCTGCATC
951 CAACAAGGAT TGCTTTGAAA AGGAAAAAGA TAGCGAAGAC TTCATAATGT
1001 GTGGTACCTG GCTCGGCAGC AGACCCTGTC CCAATGGTTC TACGTGCGAT
1051 AAAACCACAT TGAACCCAGA CAATAATTAT ACAAAGTTTG ACAACTTTGG
1101 CTGGTCCTTT CTCGCCATGT TCCGGGTTAT GACTCAAGAC TCCTGGGAGA
1151 GGCTTTACCG ACAGATCCTG CGGACCTCTG GGATCTACTT TGTCTTCTTC
1201 TTCGTGGTGG TCATCTTCCT GGGCTCCTTC TACCTGCTTA ACCTAACCTT
```

Figure 1B: SEQ ID NO:1

1251 GGCTGTTGTC ACCATGGCTT ATGAAGAACA GAACAGAAAT GTAGCTGCTG
 1301 AGACAGAGGC CAAGGAGAAA ATGTTTCAGG AAGCCCAGCA GCTGTTAAGG
 1351 GAGGAGAAGG AGGCTCTGGT TGCCATGGGA ATTGACAGAA GTTCCCTTAA
 1401 TTCCCTTCAA GCTTCATCCT TTTCCCCGAA GAAGAGGAAG TTTTTCGGTA
 1451 GTAAGACAAG AAAGTCCTTC TTTATGAGAG GGTCCAAGAC GGCCCAAGCC
 1501 TCAGCGTCTG ATTCAGAGGA CGATGCCTCT AAAAATCCAC AGCTCCTTGA
 1551 GCAGACCAAA CGACTGTCCC AGAACTTGCC AGTGGATCTC TTTGATGAGC
 1601 ACGTGGACCC CCTCCACAGG CAGAGAGCGC TGAGCGCTGT CAGTATCTTA
 1651 ACCATCACCA TGCAGGAACA AGAAAAATTC CAGGAGCCTT GTTCCCATG
 1701 TGGGAAAAAT TTGGCCTCTA AGTACCTGGT GTGGGACTGT AGCCCTCAGT
 1751 GGCTGTGCAT AAAGAAGGTC CTGCGGACCA TCATGACGGA TCCCTTTACT
 1801 GAGCTGGCCA TCACCATCTG CATCATCATC AATACCGTTT TCTTAGCCGT
 1851 GGAGCACCAC AACATGGATG ACAACTTAAA GACCATACTG AAAATAGGAA
 1901 ACTGGGTTTT CACGGGAATT TTCATAGCGG AAATGTGTCT CAAGATCATC
 1951 GCGCTCGACC CTTACCACTA CTTCCGGCAC GGCTGGAATG TTTTGTACAG
 2001 CATCGTGGCC CTCCTGAGTC TCGCTGATGT GCTCTACAAC ACACTGTCTG
 2051 ATAACAATAG GTCTTTCCTG GCTTCCCTCA GAGTGCTGAG GGTCTTCAAG
 2101 TTAGCCAAAT CCTGGCCCAC GTTAAACACT CTCATTAAGA TCATCGGCCA
 2151 CTCCGTGGGC GCGCTTGGA ACCTGACTGT GGTCTGACT ATCGTGGTCT
 2201 TCATCTTTTC TGTGGTGGGC ATGCGGCTCT TCGGCACCAA GTTTAACAAG
 2251 ACCGCCTACG CCACCCAGGA GCGGCCCAGG CGGCGCTGGC ACATGGATAA
 2301 TTTCTACCAC TCCTTCCTGG TGGTGTTCG CATCCTCTGT GGGGAATGGA
 2351 TCGAGAACAT GTGGGGCTGC ATGCAGGATA TGGACGGCTC CCCGTTGTGC
 2401 ATCATTGTCT TTGTCCTGAT AATGGTGATC GGAAGCTTG TGGTGCTTAA

Figure 1C: SEQ ID NO:1

2451 CCTCTTCATT GCCTTGCTGC TCAATTCCTT CAGCAATGAG GAGAAGGATG
2501 GGAGCCTGGA AGGAGAGACC AGGAAAACCA AAGTGCAGCT AGCCCTGGAT
2551 CGGTTCCGCC GGGCCTTCTC CTTTCATGCTG CACGCTCTTC AGAGTTTTTG
2601 TTGCAAGAAA TGCAGGAGGA AAAACTCGCC AAAGCCAAAA GAGACAACAG
2651 AAAGCTTTGC TGGTGAGAAT AAAGACTCAA TCCTCCCGGA TGCGAGGCCC
2701 TGGAAGGAGT ATGATACAGA CATGGCTTTG TACACTGGAC AGGCCGGGGC
2751 TCCGCTGGCC CCACTCGCAG AGGTAGAGGA CGATGTGGAA TATTGTGGTG
2801 AAGGCGGTGC CCTACCCACC TCACAACATA GTGCTGGAGT TCAGGCCGGT
2851 GACCTCCCTC CAGAGACCAA GCAGCTCACT AGCCCGGATG ACCAAGGGGT
2901 TGAAATGGAA GTATTTTCTG AAGAAGATCT GCATTTAAGC ATACAGAGTC
2951 CTCGAAAGAA GTCTGACGCA GTGAGCATGC TCTCGGAATG CAGCACAATT
3001 GACCTGAATG ATATCTTTAG AAATTTACAG AAAACAGTTT CCCCCAAAA
3051 GCAGCCAGAT AGATGCTTTC CCAAGGGCCT TAGTTGTCAC TTTCTATGCC
3101 ACAAACAGA CAAGAGAAAG TCCCCCTGGG TCCTGTGGTG GAACATTCGG
3151 AAAACCTGCT ACCAAATCGT GAAGCACAGC TGGTTTGAGA GTTTCATAAT
3201 CTTTGTATT CTGCTGAGCA GTGGAGCGCT GATATTTGAA GATGTCAATC
3251 TCCCCAGCCG GCCCCAAGTT GAGAAATTAC TAAGGTGTAC CGATAATATT
3301 TTCACATTTA TTTTCCTCCT GGAAATGATC CTGAAGTGGG TGGCCTTTGG
3351 ATTCCGGAGG TATTTACCA GTGCCTGGTG CTGGCTTGAT TTCCTCATTG
3401 TGGTGGTGTC TGTGCTCAGT CTCATGAATC TACCAAGCTT GAAGTCCTTC
3451 CGGACTCTGC GGGCCCTGAG ACCTCTGCGG GCGCTGTCCC AGTTTGAAGG
3501 AATGAAGGTT GTCGTCTACG CCCTGATCAG CGCCATACCT GCCATTCTCA
3551 ATGTCTTGCT GGTCTGCCTC ATTTTCTGGC TCGTATTTTG TATCTTGGGA
3601 GTAAATTTAT TTTCTGGGAA GTTTGGAAGG TGCATTAACG GGACAGACAT

Figure 1D: SEQ ID NO:1

3651 AAATATGTAT TTGGATTTTA CCGAAGTTCC GAACCGAAGC CAATGTAACA
 3701 TTAGTAATTA CTCGTGGAAG GTCCCGCAGG TCAACTTTGA CAACGTGGGG
 3751 AATGCCTATC TCGCCCTGCT GCAAGTGGCA ACCTATAAGG GCTGGCTGGA
 3801 AATCATGAAT GCTGCTGTCG ATTCCAGAGA GAAAGACGAG CAGCCGGACT
 3851 TTGAGGCGAA CCTCTACGCG TATCTCTACT TTGTGGTTTT TATCATCTTC
 3901 GGCTCCTTCT TTACCCTGAA CCTCTTTATC GGTGTTATTA TTGACAACTT
 3951 CAATCAGCAG CAGAAAAAGT TAGGTGGCCA AGACATTTTT ATGACAGAAG
 4001 AACAGAAGAA ATATTACAAT GCAATGAAAA AGTTAGGAAC CAAGAAACCT
 4051 CAAAAGCCCA TCCCAAGGCC CCTGAACAAA TGTCAAGCCT TTGTGTTCTGA
 4101 CCTGGTCACA AGCCAGGTCT TTGACGTCAT CATTCTGGGT CTTATTGTCT
 4151 TAAATATGAT TATCATGATG GCTGAATCTG CCGACCAGCC CAAAGATGTG
 4201 AAGAAAACCT TTGATATCCT CAACATAGCC TTCGTGGTCA TCTTTACCAT
 4251 AGAGTGCTC ATCAAAGTCT TTGCTTTGAG GCAACACTAC TTCACCAATG
 4301 GCTGGAACTT ATTTGATTGT GTGGTCGTGG TTCTTTCTAT CATTAGTACC
 4351 CTGGTTTCCC GCTTGGAGGA CAGTGACATT TCTTTCCCGC CCACGCTCTT
 4401 CAGAGTCGTC CGCTTGGCTC GGATTGGTCG AATCCTCAGG CTGGTCCGGG
 4451 CTGCCCCGGG AATCAGGACC CTCCTCTTTG CTTTGATGAT GTCTCTCCCC
 4501 TCTCTCTTCA ACATCGGTCT GCTGCTCTTC CTGGTGATGT TCATTTACGC
 4551 CATCTTTGGG ATGAGCTGGT TTTCCAAAGT GAAGAAGGGC TCCGGGATCG
 4601 ACGACATCTT CAACTTCGAG ACCTTTACGG GCAGCATGCT GTGCCTCTTC
 4651 CAGATAACCA CTTCGGCTGG CTGGGATACC CTCCTCAACC CCATGCTGGA
 4701 GGCAAAAGAA CACTGCAACT CCTCCTCCCA AGACAGCTGT CAGCAGCCGC
 4751 AGATAGCCGT CGTCTACTTC GTCAGTTACA TCATCATCTC CTTCTCATC
 4801 GTGGTCAACA TGTACATCGC TGTGATCCTC GAGAACTTCA ACACAGCCAC

Figure 1E: SEQ ID NO: 1

4851 GGAGGAGAGC GAGGACCCCTC TGGGAGAGGA CGACTTTGAA ATCTTCTATG
 4901 AGGTCTGGGA GAAGTTTGAC CCCGAGGCGT CGCAGTTCAT CCAGTATTCG
 4951 GCCCTCTCTG ACTTTGCGGA CGCCCTGCCG GAGCCGTTGC GTGTGGCCAA
 5001 GCCGAATAAG TTTCAGTTTC TAGTGATGGA CTTGCCCATG GTGATGGGCG
 5051 ACCGCCTCCA TTGCATGGAT GTTCTCTTTG CTTTCACTAC CAGGGTCCTC
 5101 GGGGACTCCA GCGGCTTGGA TACCATGAAA ACCATGATGG AGGAGAAGTT
 5151 TATGGAGGCC AACCCTTTTA AGAAGCTCTA CGAGCCCATA GTCACCACCA
 5201 CCAAGAGGAA GGAGGAGGAG CAAGGCGCCG CCGTCATCCA GAGGGCCTAC
 5251 CGGAAACACA TGGAGAAGAT GGTCAAACCTG AGGCTGAAGG ACAGGTCAAG
 5301 TTCATCGCAC CAGGTGTTTT GCAATGGAGA CTTGTCCAGC TTGGATGTGG
 5351 CCAAGGTCAA GGTTACAAT GACTGAACCC TCATCTCCAC CCCTACCTCA
 5401 CTGCCTCACA GCTTAGCCTC CAGCCTCTGG CGAGCAGGCG GCAGACTCAC
 5451 TGAACACAGG CCGTTCGATC TGTGTTTTTG GCTGAACGAG GTGACAGGTT
 5501 GGCCTCCATT TTTAAATGAC TCTTGGAAG ATTTTCATGTA GAGAGATGTT
 5551 AGAAGGGACT GCAAAGGACA CCGACCATAA CGGAAGGCCT GGAGGACAGT
 5601 CCAACTTACA TAAAGATGAG AAACAAGAAG GAAAGATCCC AGGAAAACCT
 5651 CAGATTGTGT TCTCAGTACA TCCCCCAATG TGTCTGTTCG GTGTTTTGAG
 5701 TATGTGACCT GCCACATGTA GCTCTTTTTT GCATGTACGT CAAAACCCCTG
 5751 CAGTAAGTTG ATAGCTTGCT ACGGGTGTTT CTACCAGCAT CACAGAATTG
 5801 GGTGTATGAC TCAAACCTAA AAGCATGACT CTGACTTGTC AGTCAGCACC
 5851 CCGACTTTCA GACGCTCCAA TCTCTGTCCC AGGTGTCTAA CGAATAAATA
 5901 GGTAAAAG

Figure 2A: SEQ ID NO: 2

Met Glu Glu Arg Tyr Tyr Pro Val Ile Phe Pro Asp Glu Arg Asn Phe
 1 5 10 15
 Arg Pro Phe Thr Ser Asp Ser Leu Ala Ala Ile Glu Lys Arg Ile Ala
 20 25 30
 Ile Gln Lys Glu Arg Lys Lys Ser Lys Asp Lys Ala Ala Ala Glu Pro
 35 40 45
 Gln Pro Arg Pro Gln Leu Asp Leu Lys Ala Ser Arg Lys Leu Pro Lys
 50 55 60
 Leu Tyr Gly Asp Ile Pro Pro Glu Leu Val Ala Lys Pro Leu Glu Asp
 65 70 75 80
 Leu Asp Pro Phe Tyr Lys Asp His Lys Thr Phe Met Val Leu Asn Lys
 85 90 95
 Lys Arg Thr Ile Tyr Arg Phe Ser Ala Lys Arg Ala Leu Phe Ile Leu
 100 105 110
 Gly Pro Phe Asn Pro Leu Arg Ser Leu Met Ile Arg Ile Ser Val His
 115 120 125
 Ser Val Phe Ser Met Phe Ile Ile Cys Thr Val Ile Ile Asn Cys Met
 130 135 140
 Phe Met Ala Asn Ser Met Glu Arg Ser Phe Asp Asn Asp Ile Pro Glu
 145 150 155 160
 Tyr Val Phe Ile Gly Ile Tyr Ile Leu Glu Ala Val Ile Lys Ile Leu
 165 170 175
 Ala Arg Gly Phe Ile Val Asp Glu Phe Ser Phe Leu Arg Asp Pro Trp
 180 185 190
 Asn Trp Leu Asp Phe Ile Val Ile Gly Thr Ala Ile Ala Thr Cys Phe
 195 200 205
 Pro Gly Ser Gln Val Asn Leu Ser Ala Leu Arg Thr Phe Arg Val Phe
 210 215 220
 Arg Ala Leu Lys Ala Ile Ser Val Ile Ser Gly Leu Lys Val Ile Val
 225 230 235 240
 Gly Ala Leu Leu Arg Ser Val Lys Lys Leu Val Asp Val Met Val Leu
 245 250 255
 Thr Leu Phe Cys Leu Ser Ile Phe Ala Leu Val Gly Gln Gln Leu Phe
 260 265 270
 Met Gly Ile Leu Asn Gln Lys Cys Ile Lys His Asn Cys Gly Pro Asn
 275 280 285

7/27

Pro Ala Ser Asn Lys Asp Cys Phe Glu Lys Glu Lys Asp Ser Glu Asp
290 295 300
Phe Ile Met Cys Gly Thr Trp Leu Gly Ser Arg Pro Cys Pro Asn Gly
305 310 315 320

Figure 2B: SEQ ID NO: 2

Ser Thr Cys Asp Lys Thr Thr Leu Asn Pro Asp Asn Asn Tyr Thr Lys			
325	330	335	
Phe Asp Asn Phe Gly Trp Ser Phe Leu Ala Met Phe Arg Val Met Thr			
340	345	350	
Gln Asp Ser Trp Glu Arg Leu Tyr Arg Gln Ile Leu Arg Thr Ser Gly			
355	360	365	
Ile Tyr Phe Val Phe Phe Phe Val Val Val Ile Phe Leu Gly Ser Phe			
370	375	380	
Tyr Leu Leu Asn Leu Thr Leu Ala Val Val Thr Met Ala Tyr Glu Glu			
385	390	395	400
Gln Asn Arg Asn Val Ala Ala Glu Thr Glu Ala Lys Glu Lys Met Phe			
405	410	415	
Gln Glu Ala Gln Gln Leu Leu Arg Glu Glu Lys Glu Ala Leu Val Ala			
420	425	430	
Met Gly Ile Asp Arg Ser Ser Leu Asn Ser Leu Gln Ala Ser Ser Phe			
435	440	445	
Ser Pro Lys Lys Arg Lys Phe Phe Gly Ser Lys Thr Arg Lys Ser Phe			
450	455	460	
Phe Met Arg Gly Ser Lys Thr Ala Gln Ala Ser Ala Ser Asp Ser Glu			
465	470	475	480
Asp Asp Ala Ser Lys Asn Pro Gln Leu Leu Glu Gln Thr Lys Arg Leu			
485	490	495	
Ser Gln Asn Leu Pro Val Asp Leu Phe Asp Glu His Val Asp Pro Leu			
500	505	510	
His Arg Gln Arg Ala Leu Ser Ala Val Ser Ile Leu Thr Ile Thr Met			
515	520	525	
Gln Glu Gln Glu Lys Phe Gln Glu Pro Cys Phe Pro Cys Gly Lys Asn			
530	535	540	
Leu Ala Ser Lys Tyr Leu Val Trp Asp Cys Ser Pro Gln Trp Leu Cys			
545	550	555	560
Ile Lys Lys Val Leu Arg Thr Ile Met Thr Asp Pro Phe Thr Glu Leu			
565	570	575	
Ala Ile Thr Ile Cys Ile Ile Ile Asn Thr Val Phe Leu Ala Val Glu			
580	585	590	
His His Asn Met Asp Asp Asn Leu Lys Thr Ile Leu Lys Ile Gly Asn			

9/27

595	600	605
Trp Val Phe Thr Gly Ile Phe Ile Ala Glu Met Cys Leu Lys Ile Ile		
610	615	620

Figure 2C: SEQ ID NO: 2

Ala	Leu	Asp	Pro	Tyr	His	Tyr	Phe	Arg	His	Gly	Trp	Asn	Val	Phe	Asp
625						630				635					640
Ser	Ile	Val	Ala	Leu	Leu	Ser	Leu	Ala	Asp	Val	Leu	Tyr	Asn	Thr	Leu
			645						650					655	
Ser	Asp	Asn	Asn	Arg	Ser	Phe	Leu	Ala	Ser	Leu	Arg	Val	Leu	Arg	Val
			660					665					670		
Phe	Lys	Leu	Ala	Lys	Ser	Trp	Pro	Thr	Leu	Asn	Thr	Leu	Ile	Lys	Ile
		675					680						685		
Ile	Gly	His	Ser	Val	Gly	Ala	Leu	Gly	Asn	Leu	Thr	Val	Val	Leu	Thr
	690					695					700				
Ile	Val	Val	Phe	Ile	Phe	Ser	Val	Val	Gly	Met	Arg	Leu	Phe	Gly	Thr
705				710					715					720	
Lys	Phe	Asn	Lys	Thr	Ala	Tyr	Ala	Thr	Gln	Glu	Arg	Pro	Arg	Arg	Arg
			725					730					735		
Trp	His	Met	Asp	Asn	Phe	Tyr	His	Ser	Phe	Leu	Val	Val	Phe	Arg	Ile
		740					745						750		
Leu	Cys	Gly	Glu	Trp	Ile	Glu	Asn	Met	Trp	Gly	Cys	Met	Gln	Asp	Met
	755					760						765			
Asp	Gly	Ser	Pro	Leu	Cys	Ile	Ile	Val	Phe	Val	Leu	Ile	Met	Val	Ile
	770				775							780			
Gly	Lys	Leu	Val	Val	Leu	Asn	Leu	Phe	Ile	Ala	Leu	Leu	Leu	Asn	Ser
785			790						795					800	
Phe	Ser	Asn	Glu	Glu	Lys	Asp	Gly	Ser	Leu	Glu	Gly	Glu	Thr	Arg	Lys
			805					810					815		
Thr	Lys	Val	Gln	Leu	Ala	Leu	Asp	Arg	Phe	Arg	Arg	Ala	Phe	Ser	Phe
		820					825						830		
Met	Leu	His	Ala	Leu	Gln	Ser	Phe	Cys	Cys	Lys	Lys	Cys	Arg	Arg	Lys
	835					840						845			
Asn	Ser	Pro	Lys	Pro	Lys	Glu	Thr	Thr	Glu	Ser	Phe	Ala	Gly	Glu	Asn
	850				855						860				
Lys	Asp	Ser	Ile	Leu	Pro	Asp	Ala	Arg	Pro	Trp	Lys	Glu	Tyr	Asp	Thr
865			870					875					880		
Asp	Met	Ala	Leu	Tyr	Thr	Gly	Gln	Ala	Gly	Ala	Pro	Leu	Ala	Pro	Leu
			885					890					895		

11/27

Ala Glu Val Glu Asp Asp Val Glu Tyr Cys Gly Glu Gly Gly Ala Leu
900 905 910

Figure 2D: SEQ ID NO: 2

Pro Thr Ser Gln His Ser Ala Gly Val Gln Ala Gly Asp Leu Pro Pro
 915 920 925
 Glu Thr Lys Gln Leu Thr Ser Pro Asp Asp Gln Gly Val Glu Met Glu
 930 935 940
 Val Phe Ser Glu Glu Asp Leu His Leu Ser Ile Gln Ser Pro Arg Lys
 945 950 955 960
 Lys Ser Asp Ala Val Ser Met Leu Ser Glu Cys Ser Thr Ile Asp Leu
 965 970 975
 Asn Asp Ile Phe Arg Asn Leu Gln Lys Thr Val Ser Pro Lys Lys Gln
 980 985 990
 Pro Asp Arg Cys Phe Pro Lys Gly Leu Ser Cys His Phe Leu Cys His
 995 1000 1005
 Lys Thr Asp Lys Arg Lys Ser Pro Trp Val Leu Trp Trp Asn Ile Arg
 1010 1015 1020
 Lys Thr Cys Tyr Gln Ile Val Lys His Ser Trp Phe Glu Ser Phe Ile
 1025 1030 1035 1040
 Ile Phe Val Ile Leu Leu Ser Ser Gly Ala Leu Ile Phe Glu Asp Val
 1045 1050 1055
 Asn Leu Pro Ser Arg Pro Gln Val Glu Lys Leu Leu Arg Cys Thr Asp
 1060 1065 1070
 Asn Ile Phe Thr Phe Ile Phe Leu Leu Glu Met Ile Leu Lys Trp Val
 1075 1080 1085
 Ala Phe Gly Phe Arg Arg Tyr Phe Thr Ser Ala Trp Cys Trp Leu Asp
 1090 1095 1100
 Phe Leu Ile Val Val Val Ser Val Leu Ser Leu Met Asn Leu Pro Ser
 1105 1110 1115 1120
 Leu Lys Ser Phe Arg Thr Leu Arg Ala Leu Arg Pro Leu Arg Ala Leu
 1125 1130 1135
 Ser Gln Phe Glu Gly Met Lys Val Val Val Tyr Ala Leu Ile Ser Ala
 1140 1145 1150
 Ile Pro Ala Ile Leu Asn Val Leu Leu Val Cys Leu Ile Phe Trp Leu
 1155 1160 1165
 Val Phe Cys Ile Leu Gly Val Asn Leu Phe Ser Gly Lys Phe Gly Arg
 1170 1175 1180
 Cys Ile Asn Gly Thr Asp Ile Asn Met Tyr Leu Asp Phe Thr Glu Val

13 | 27

1185

1190

1195

1200

:

Pro Asn Arg Ser Gln Cys Asn Ile Ser Asn Tyr Ser Trp Lys Val Pro

1205

1210

1215

Figure 2E: SEQ ID NO: 2

Gln Val Asn Phe Asp Asn Val Gly Asn Ala Tyr Leu Ala Leu Leu Gln
 1220 1225 1230
 Val Ala Thr Tyr Lys Gly Trp Leu Glu Ile Met Asn Ala Ala Val Asp
 1235 1240 1245
 Ser Arg Glu Lys Asp Glu Gln Pro Asp Phe Glu Ala Asn Leu Tyr Ala
 1250 1255 1260
 Tyr Leu Tyr Phe Val Val Phe Ile Ile Phe Gly Ser Phe Phe Thr Leu
 1265 1270 1275 1280
 Asn Leu Phe Ile Gly Val Ile Ile Asp Asn Phe Asn Gln Gln Gln Lys
 1285 1290 1295
 Lys Leu Gly Gly Gln Asp Ile Phe Met Thr Glu Glu Gln Lys Lys Tyr
 1300 1305 1310
 Tyr Asn Ala Met Lys Lys Leu Gly Thr Lys Lys Pro Gln Lys Pro Ile
 1315 1320 1325
 Pro Arg Pro Leu Asn Lys Cys Gln Ala Phe Val Phe Asp Leu Val Thr
 1330 1335 1340
 Ser Gln Val Phe Asp Val Ile Ile Leu Gly Leu Ile Val Leu Asn Met
 1345 1350 1355 1360
 Ile Ile Met Met Ala Glu Ser Ala Asp Gln Pro Lys Asp Val Lys Lys
 1365 1370 1375
 Thr Phe Asp Ile Leu Asn Ile Ala Phe Val Val Ile Phe Thr Ile Glu
 1380 1385 1390
 Cys Leu Ile Lys Val Phe Ala Leu Arg Gln His Tyr Phe Thr Asn Gly
 1395 1400 1405
 Trp Asn Leu Phe Asp Cys Val Val Val Val Leu Ser Ile Ile Ser Thr
 1410 1415 1420
 Leu Val Ser Arg Leu Glu Asp Ser Asp Ile Ser Phe Pro Pro Thr Leu
 1425 1430 1435 1440
 Phe Arg Val Val Arg Leu Ala Arg Ile Gly Arg Ile Leu Arg Leu Val
 1445 1450 1455
 Arg Ala Ala Arg Gly Ile Arg Thr Leu Leu Phe Ala Leu Met Met Ser
 1460 1465 1470
 Leu Pro Ser Leu Phe Asn Ile Gly Leu Leu Leu Phe Leu Val Met Phe
 1475 1480 1485
 Ile Tyr Ala Ile Phe Gly Met Ser Trp Phe Ser Lys Val Lys Lys Gly

1490 1495 1500
Ser Gly Ile Asp Asp Ile Phe Asn Phe Glu Thr Phe Thr Gly Ser Met
1505 1510 1515 1520

Figure 2F: SEQ ID NO: 2

Leu Cys Leu Phe Gln Ile Thr Thr Ser Ala Gly Trp Asp Thr Leu Leu
 1525 1530 1535
 Asn Pro Met Leu Glu Ala Lys Glu His Cys Asn Ser Ser Ser Gln Asp
 1540 1545 1550
 Ser Cys Gln Gln Pro Gln Ile Ala Val Val Tyr Phe Val Ser Tyr Ile
 1555 1560 1565
 Ile Ile Ser Phe Leu Ile Val Val Asn Met Tyr Ile Ala Val Ile Leu
 1570 1575 1580
 Glu Asn Phe Asn Thr Ala Thr Glu Glu Ser Glu Asp Pro Leu Gly Glu
 1585 1590 1595 1600
 Asp Asp Phe Glu Ile Phe Tyr Glu Val Trp Glu Lys Phe Asp Pro Glu
 1605 1610 1615
 Ala Ser Gln Phe Ile Gln Tyr Ser Ala Leu Ser Asp Phe Ala Asp Ala
 1620 1625 1630
 Leu Pro Glu Pro Leu Arg Val Ala Lys Pro Asn Lys Phe Gln Phe Leu
 1635 1640 1645
 Val Met Asp Leu Pro Met Val Met Gly Asp Arg Leu His Cys Met Asp
 1650 1655 1660
 Val Leu Phe Ala Phe Thr Thr Arg Val Leu Gly Asp Ser Ser Gly Leu
 1665 1670 1675 1680
 Asp Thr Met Lys Thr Met Met Glu Glu Lys Phe Met Glu Ala Asn Pro
 1685 1690 1695
 Phe Lys Lys Leu Tyr Glu Pro Ile Val Thr Thr Thr Lys Arg Lys Glu
 1700 1705 1710
 Glu Glu Gln Gly Ala Ala Val Ile Gln Arg Ala Tyr Arg Lys His Met
 1715 1720 1725
 Glu Lys Met Val Lys Leu Arg Leu Lys Asp Arg Ser Ser Ser Ser His
 1730 1735 1740
 Gln Val Phe Cys Asn Gly Asp Leu Ser Ser Leu Asp Val Ala Lys Val
 1745 1750 1755 1760
 Lys Val His Asn Asp
 1765

Figure 2G: SEQ ID NO:2

```

1  MEERYYPVIF PDERNFRPPT SDSLAAIEKR IAIQKERKKS KDKAAAEPQP
51  RPQLDLKASR KLPKLYGDIP PELVAKPLED LDPFYKDHKT FMVLNKKRTI
101 YRFSAKRALF ILGPFNPLRS LMIRISVHSV FSMFIICTVI INCMFMANSM
    |-----IS1-----|
151 ERSFDNDIPE YVFIGIYILE AVIKILARGF IVDEFSFLRD PWNWLD FIVI
    |-----IS2-----| |-----IS3-----|
201 GTAIATCFPG SQVNLSALRT FRVFRALKAI SVISGLKVIV GALLRSVKKL
    -----| • |-----IS4-----|
251 VDVMTLFC LSIFALVGQQ LFMGILNQKC IKHNCGP NPA SNKDCFEKEK
    |-----IS5-----|
301 DSEDFIMCGT WLGSRPCNG STCDKTTLNP DNNTYTKFDNF GWSFLAMFRV
    • •
351 MTQDSWERLY RQILRTSGIY FVFFV VVIF LGSFYLLNLT LAVVTMAYEE
    • |-----IS6-----|
401 QNRNVAAETE AKEKMFQEAQ QLLREEKEAL VAMGIDRSSL NSLQASSFSP
451 KKRKFFGSKT RKSFFMRGSK TAQASASDSE DDASKNPQLL EQTKRLSQNL
    •
501 PVDLFD EHV D PLHRQRALSA VSILTITMQE QEKFQEP CFP CGKNLASKYL
551 VWDCSPQWLC IKKVLRTIMT DPFTELAITI CIIINTVFLA VEHNMDDNL
    |-----IIS1-----|
601 KTIKIGNWV FTGIFIAEMC LKIIALDPYH YFRHGWNVFD SIVALLSLAD
    |-----IIS2-----| |-----IIS3-----|
651 VLYNTLSDNN RSFLASLRVL RVFKLAKSWP TLNTLIKIIG HSVGALGNLT
    ----| • |-----IIS4-----| •|
701 VVLTIVVFIF SVVGMR LFGT KFNKTAYATQ ERPRRRWHMD NFYHSFLVVF
    -----IIS5-----| •
751 RILCGEWIEN MWGCMQMDG SPLCIIVFVL IMVIGKLVVL NLFIALLLNS
    |-----IIS6-----|
801 FSNEEKD GSL EGETRKT KVQ LALDRFRRAF SFMLHALQSF CCKKCRKNS
    •
851 PKPKETTESF AGENKDSILP DARFWKEYDT DMALYTGQAG APLAPLAEVE
901 DDVEYCGEGG ALPTSQHSAG VQAGDLPPET KQLTSPDDQG VEMEVFSEED
951 LHLSIQSPRK KSDAVSMLSE CSTIDLNDIF RNLQKTVSPK KQPDRCFPKG
    •
1001 LSCHFLCHKT DKRKSPWWLW WNIRKTCYQI VKHSWFESFI IFVILLSSGA
    |-----IIIS1-----|
1051 LIFEDVN LPS RPQVEKLLRC TDNIFTFIFL LEMILKWVAF GFRRYFTSAW
    --| |-----IIIS2-----| |
1101 CWLDFLIVV SVLSLMNLPS LKSFR LRAL RPLRALSQFE GMKV VVYALI
    ----IIIS3-----| |-----IIIS4-----|
1151 SAIPAILNLV LVCLIFWLVF CILGVNLFSG KFGRCINGTD INMYLDFTEV
    |-----IIIS5-----| •
1201 PNRSQCNISN YSWKVPQVNF DNVGNAYLAL LQVATYKGWL EIMNAAVDSR
    • • •
1251 EKDEQPD FEA NLYAYLYFVV FIIFGSFFTL NLFIVGIIDN FNQQQKKLGG
    |-----IIIS6-----|

```

Figure 2H: SEQ ID NO: 2

```

1301 QDIFMTEEQK KYYNAMKKG TKKPQKPIPR PLNKCQAFVF DLVTSQVFDV
                                     |-----
1351 IILGLIVLNM IIMMAESADQ PKDVKKTFDI LNIAFVVIFT IECLIKVFAL
IVS1-----|-----IVS2-----|
1401 RQHYFTNGWN LFDCVVVLS IISTLVSRLE DSDISFPPTL FRVVRLARIG
      |-----IVS3-----|-----
1451 RILRLVRAAR GIRTLLFALM MSLPSLFNIG LLLFLVMFIY AIFGMSWFSK
IVS4-----|-----IVS5-----
1501 VKKGS GIDDI FNFETFTGSM LCLFQITTS A GWDTLNPM L EAKEHCNSSS
      | O
1551 QDSCQQPQIA VVYFVSYIII SFLIVNMYI AVILENFNTA TEESEDPLGE
      |-----IVS6-----|
1601 DDFEIFYEVW EKFDPEASQF IQYSALS DFA DALPEPLRVA KPNKFQFLVM
1651 DLPMVMGDRL HCM DVLFAFT TRVLGDSSGL DTMKTMMEEK FMEANPFKKL
1701 YEPIVTTTKR KEEEQGA AVI QRAYRKHMEK MVKLSLKDRS SSSHQVFCNG
1751 DLSSLDVAKV KVHND*

```

Figure 3A: SEQ ID NO:3

1 GCTGAGCAGT GGGGCACTGA TATTTGAAGA TGTTACCTT GAGAACCAAC
51 CCAAAATCCA AGAATTACTA AATTGTACTG ACATTATTTT TACACATATT
101 TTTATCCTGG AGATGGTACT AAAATGGGTA GCCTTCGGAT TTGGAAAGTA
151 TTTCAACCAGT GCCTGGTGCT GCCTTGATTT CATCATTGTG ATTGTCTCTG
201 TGACCACCCT CATTAACCTA ATGGAATTGA AGTCCTTCCG GACTCTACGA
251 GCACTGAGGC CTCTTCGTGC GCTGTCCCAG TTTGAAGGAA TGAAGGTGGT
301 GGTCAATGCT CTCATAGGTG CCATACCTGC CATTCTGAAT GTTTTGCTTG
351 TCTGCCTCAT TTTCTGGCTC GTATTTTGTA TTCTGGGAGT ATACTTCTTT
401 TCTGGAAAAT TTGGGAAATG CATTAATGGA ACAGACTCAG TTATAAATTA
451 TACCATCATT ACAAATAAAA GTCAATGTGA AAGTGGCAAT TTCTCTTGGA
501 TCAACCAGAA AGTCAACTTT GACAATGTGG GAAATGCTTA CCTCGCTCTG
551 CTGCAAGTGG CAACATTTAA GGGCTGGATG GATATTATAT ATGCAGCTGT
601 TGATTCCACA GAGAAAGAAC AACAGCCAGA GTTTGAGAGC AATTCACCTG
651 GTTACATTTA CTTCGTAGTC TTTATCATCT TTGGCTCATT CTTCACTCTG
701 AATCTCTTCA TTGGCGTTAT CATTGACAAC TTCAACCAAC AGCAGAAAAA
751 GTTAGGTGGC CAAGACATTT TTATGACAGA AGAACAGAAG AAATACTATA
801 ATGCAATGAA AAAATTAGGA TCCAAAAAAC CTCAAAAACC CATTCCACGG
851 CCCGTT

Figure 4: SEQ ID NO:4

```
1   CTCAACATGG TTACGATGAT GGTGGAGACC GACGAGCAGG GCGAGGAGAA
51  GACGAAGGTT CTGGGCAGAA TCAACCAGTT CTTTGTGGCC GTCTTCACGG
101 GCGAGTGTGT GATGAAGATG TTCGCCCTGC GACAGTACTA TTTCACCAAC
151 GGCTGGAACG TGTTCGACTT CATAGTGGTG ATCCTGTCCA TTGGGAGTCT
201 GCTGTTTCT  GCAATCCTTA AGTCACTGGA AAACACTTTC TCCCCGACGC
251 TCTTCCGGGT CATCCGTCTG GCCAGGATCG GCCGCATCCT CAGGCTGATC
301 CGAGCAGCCA AGGGGATTCG CACGCTGCTC TTCGCCCTCA TGATGTCCCT
351 GCCCCGCCCTC TTCAACATCG GCCTCCTCCT CTTCTCTCGtC ATGTTTCATCT
401 ACTCCATCTT CGGCATGGCC AGCTTCGCTA ACGTCGTGGA CGAGGCCGGC
451 ATCGACGACA TGTTCAACTT CAAGACCTTT GGCAACAGCA TGCTGTGCCT
501 GTTCCAGATC ACCACCTCGG CCGGCTGGGA CGGCCTCCTC AGCCCCATCC
551 TCAACACGGG GCCTCCCTAC TGCGACCCCA ACCTGCCCAA CAGCAACGGC
601 TCCCGGGGGA ACTGCGGGAG CCCGGCGGTG GGCATCATCT TCTTCACCAC
651 CTACATCATC ATCTCCTTCC TCATCGTGGT CAACATGTAT ATCGCAGTCA
701 TC
```

Figure 5A: SEQ ID NO: 5

```

1  GTCGACTCTA GATCAGGGTG AAGATGGAGG AGAGGTACTA CCCGGTGATC
51  TTCCCGGACG AGCGGAATTT CCGCCCCTTC ACTTCCGACT CTCTGGCTGC
101 CATAGAGAAG CGGATTGCTA TCCAAAAGGA GAGGAAGAAG TCCAAAGACA
151 AGGCGGCAGC TGAGCCCCAG CCTCGGCCTC AGCTTGACCT AAAGGCCTCC
201 AGGAAGTTAC CTAAGCTTTA TGGTGACATT CCCCTGAGC TTGTAGCGAA
251 GCCTCTGGAA GACCTGGACC CATTCTACAA AGACCATAAG ACATTCATGG
301 TGTGTAACAA GAAGAGAACA ATTTATCGCT TCAGCGCCAA GCGGGCCTTG
351 TTCATTCTGG GGCCTTTTAA TCCCCTCAGA AGCTTAATGA TTCGTATCTC
401 TGTCCATTCA GTCTTTAGCA TGTTATCAT CTGCACGGTG ATCATCAACT
451 GTATGTTTCA GGCGAATTCT ATGGAGAGAA GTTTCGACAA CGACATTCCC
501 GAATACGTCT TCATTGGGAT TTATATTTTA GAAGCTGTGA TTAAAATATT
551 GGCAAGAGGC TTCATTGTGG ATGAGTTTTT CTTCCTCCGA GATCCGTGGA
601 ACTGGCTGGA CTTCAATTGTC ATTGGAACAG CGATCGCAAC TTGTTTTCCG
651 GGCAGCCAAG TCAATCTTTC AGCTCTTCGT ACCTTCCGAG TGTTTCAGAGC
701 TCTGAAGGCG ATTTCAGTTA TCTCAGGTCT GAAGGTCATC GTAGGTGCCC
751 TGCTGCGCTC GGTGAAGAAG CTGGTAGACG TGATGGTCCT CACTCTCTTC
801 TGCCTCAGCA TCTTTGCCCT GGTCGGTCAG CAGCTGTTCA TGGGAATTCT
851 GAACCAGAAG TGTATTAAGC ACAACTGTGG CCCCAACCCT GCATCCAACA
901 AGGATTGCTT TGAAAAGGAA AAAGATAGCG AAGACTTCAT AATGTGTGGT
951 ACCTGGCTCG GCAGCAGACC CTGTCCCAAT GGTTCTACGT GCGATAAAAC
1001 CACATTGAAC CCAGACAATA ATTATACAAA GTTTGACAAC TTTGGCTGGT
1051 CCTTTCTCGC CATGTTCCGG GTTATGACTC AAGACTCCTG GGAGAGGCTT
1101 TACCGACAGA TCCTGCGGAC CTCTGGGATC TACTTTGTCT TCTTCTTCGT

```

Figure 5B: SEQ ID NO: 5

1151 GGTGGTCATC TTCCTGGGCT CCTTCTACCT GCTTAACCTA ACCCTGGCTG
1201 TTGTCACCAT GGCTTATGAA GAACAGAACA GAAATGTAGC TGCTGAGACA
1251 GAGGCCAAGG AGAAAATGTT TCAGGAAGCC CAGCAGCTGT TAAGGGAGGA
1301 GAAGGAGGCT CTGGTTGCCA TGGAATTGA CAGAAGTTCC CTTAATTCCC
1351 TTCAAGCTTC ATCCTTTTCC CCGAAGAAGA GGAAGTTTTT CGGTAGTAAG
1401 ACAAGAAAGT CCTTCTTTAT GAGAGGGTCC AAGACGGCCC AAGCCTCAGC
1451 GTCTGATTCA GAGGACGATG CCTCTAAAAA TCCACAGCTC CTTGAGCAGA
1501 CCAAACGACT GTCCCAGAAC TTGCCAGTGG ATCTCTTTGA TGAGCACGTG
1551 GACCCCTCC ACAGGCAGAG AGCGCTGAGC GCTGTCAGTA TCTTAACCAT
1601 CACCATGCAG GAACAAGAAA AATTCCAGGA GCCTTGTTTC CCATGTGGGA
1651 AAAATTTGGC CTCTAAGTAC CTGGTGTGGG ACTGTAGCCC TCAGTGGCTG
1701 TGCATAAAGA AGGTCCTGCG GACCATCATG ACGGATCCCT TTACTGAGCT
1751 GGCCATCACC ATCTGCATCA TCATCAATAC CGTTTTCTTA GCCGTGGAGC
1801 ACCACAACAT GGATGACAAC TTAAAGACCA TACTGAAAAT AGGAAACTGG
1851 GTTTTACGG GAATTTTCAT AGCGGAAATG TGTCTCAAGA TCATCGCGCT
1901 CGACCCTTAC CACTACTTCC GGCACGGCTG GAATGTTTTT GACAGCATCG
1951 TGGCCCTCCT GAGTCTCGCT GATGTGCTCT ACAACACACT GTCTGATAAC
2001 AATAGGTCTT TCTTGGCTTC CCTCAGAGTG CTGAGGGTCT TCAAGTTAGC
2051 CAAATCCTGG CCCACGTTAA AACTCTCAT TAAGATCATC GGCCACTCCG
2101 TGGGCGCGCT TGGAAACCTG ACTGTGGTCC TGAATATCGT GGTCTTCATC
2151 TTTTCTGTGG TGGGCATGCG GCTCTTCGGC ACCAAGTTTA ACAAGACCGC
2201 CTACGCCACC CAGGAGCGGC CCAGGCGGCG CTGGCACATG GATAATTTCT
2251 ACCACTCCTT CCTGGTGGTG TTCCGCATCC TCTGTGGGGA ATGGATCGAG
2301 AACATGTGGG GCTGCATGCA GGATATGGAC GGCTCCCCGT TGTGCATCAT

Figure 5C: SEQ ID NO: 5

2351 TGTCTTTGTC CTGATAATGG TGATCGGGAA GCTTGTGGTG CTTAACCTCT
2401 TCATTGCCTT GCTGCTCAAT TCCTTCAGCA ATGAGGAGAA GGATGGGAGC
2451 CTGGAAGGAG AGACCAGGAA AACCAAAGTG CAGCTAGCCC TGGATCGGTT
2501 CCGCCGGGCC TTCTCCTTCA TGCTGCACGC TCTTCAGAGT TTTTGTGCA
2551 AGAAATGCAG GAGGAAAAAC TCGCCAAAGC CAAAAGAGAC AACAGAAAGC
2601 TTTGCTGGTG AGAATAAAGA CTCAATCCTC CCGGATGCGA GGCCCTGGAA
2651 GGAGTATGAT ACAGACATGG CTTTGTACAC TGGACAGGCC GGGGCTCCGC
2701 TGGCCCCACT CGCAGAGGTA GAGGACGATG TGGAAATATTG TGGTGAAGGC
2751 GGTGCCCTAC CCACCTCACA ACATAGTGCT GGAGTTCAGG CCGGTGACCT
2801 CCCTCCAGAG ACCAAGCAGC TACTAGCCC GGATGACCAA GGGGTGAAA
2851 TGGAAGTATT TTCTGAAGAA GATCTGCATT TAAGCATACA GAGTCCTCGA
2901 AAGAAGTCTG ACGCAGTGAG CATGCTCTCG GAATGCAGCA CAATTGACCT
2951 GAATGATATC TTTAGAAATT TACAGAAAAC AGTTTCCCCC AAAAAGCAGC
3001 CAGATAGATG CTTTCCCAAG GGCCTTAGTT GTCACTTTCT ATGCCACAAA
3051 ACAGACAAGA GAAAGTCCCC CTGGGTCTCG TGGTGGAACA TTCGAAAAC
3101 CTGCTACCAA ATCGTGAAGC ACAGCTGGTT TGAGAGTTTC ATAATCTTTG
3151 TTATTCTGCT GAGCAGTGGA GCGCTGATAT TTGAAGATGT CAATCTCCCC
3201 AGCCGGCCCC AAGTTGAGAA ATTACTAAGG TGTACCGATA ATATTTTCAC
3251 ATTTATTTTC CTCCTGGAAA TGATCCTGAA GTGGGTGGCC TTTGGATTCC
3301 GGAGGTATTT CACCAGTGCC TGGTGCTGGC TTGATTTCTT CATTGTGGTG
2251 GTGTCTGTGC TCAGTCTCAT GAATCTACCA AGCTTGAAGT CCTTCCGGAC
3401 TCTGCGGGCC CTGAGACCTC TCGGGCGCT GTCCCAGTTT GAAGGAATGA
3451 AGGTTGTCGT CTACGCCCTG ATCAGCGCCA TACCTGCCAT TCTCAATGTC
3501 TTGCTGGTCT GCCTCATTTT CTGGCTCGTA TTTTGTATCT TGGGAGTAAA

Figure 5D: SEQ ID NO: 5

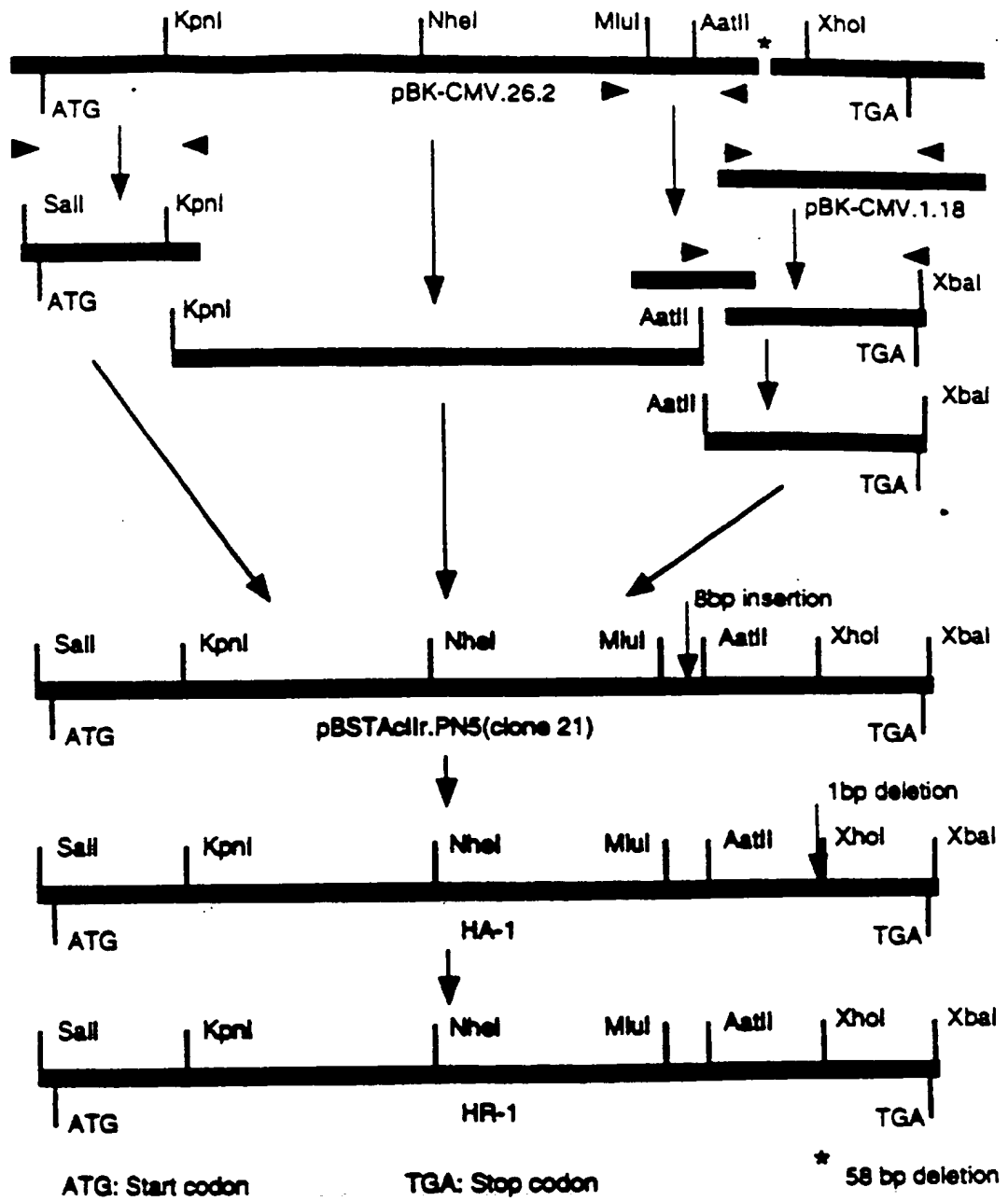
3551 TTTATTTTCT GGGAAGTTTG GAAGGTGCAT TAACGGGACA GACATAAATA
3601 TGTATTTGGA TTTTACCGAA GTTCCGAACC GAAGCCAATG TAACATTAGT
3651 AATTACTCGT GGAAGGTCCC GCAGGTCAAC TTTGACAACG TGGGGAATGC
3701 CTATCTCGCC CTGCTGCAAG TGGCAACCTA TAAGGGCTGG CTGGAAATCA
3751 TGAATGCTGC TGTGATTCC AGAGAGAAAG ACGAGCAGCC GGACTTTGAG
3801 GCGAACCTCT ACGCGTATCT CTACTTTGTG GTTTTTATCA TCTTCGGCTC
3851 CTTCTTTACC CTGAACCTCT TTATCGGTGT TATTATTGAC AACTTCAATC
3901 AGCAGCAGAA AAAGTTAGGT GGCCAAGACA TCTTCATGAC TGAGGAGCAG
3951 AAGAAATATT ACAATGCAAT GAAAAAGTTA GGAACCAAGA AACCTCAAAA
4001 GCCCATCCCA AGGCCCTGA ACAAATGTCA AGCCTTTGTG TTCGACCTGG
4051 TCACAAGCCA GGTCTTTGAC GTCATCATTC TGGGTCTTAT TGTCTTAAAT
4101 ATGATTATCA TGATGGCTGA ATCTGCCGAC CAGCCCAAAG ATGTGAAGAA
4151 AACCTTTGAT ATCCTCAACA TAGCCTTCGT GGTCATCTTT ACCATAGAGT
4201 GTCTCATCAA AGTCTTTGCT TTGAGGCAAC ACTACTTCAC CAATGGCTGG
4251 AACTTATTTG ATTGTGTGGT CGTGGTTCTT TCTATCATTA GTACCCTGGT
4301 TTCCCGCTTG GAGGACAGTG ACATTTCTTT CCCGCCACG CTCTTCAGAG
4351 TCGTCCGCTT GGCTCGGATT GGTCGAATCC TCAGGCTGGT CCGGGCTGCC
4401 CGGGGAATCA GGACCTCCT CTTTGCTTTG ATGATGTCTC TCCCCTCTCT
4451 CTTCAACATC GGTCTGCTGC TCTTCCTGGT GATGTTTATT TACGCCATCT
4501 TTGGGATGAG CTGGTTTTCC AAAGTGAAGA AGGGCTCCGG GATCGACGAC
4551 ATCTTCAACT TCGAGACCTT TACGGGCAGC ATGCTGTGCC TCTTCCAGAT
4601 AACCACTTCG GCTGGCTGGG ATACCCTCCT CAACCCCATG CTGGAGGCAA
4651 AAGAACACTG CAACTCCTCC TCCCAAGACA GCTGTCAGCA GCCGCAGATA
4701 GCCGTCGTCT ACTTCGTGAG TTACATCATC ATCTCCTTCC TCATCGTGGT

Figure 5E: SEQ ID NO: 5

4751 CAACATGTAC ATCGCTGTGA TCCTCGAGAA CTTCAACACA GCCACGGAGG
4801 AGAGCGAGGA CCCTCTGGGA GAGGACGACT TTGAAATCTT CTATGAGGTC
4851 TGGGAGAAGT TTGACCCCGA GCGGTCGCAG TTCATCCAGT ATTTCGGCCCT
4901 CTCTGACTTT GCGGACGCCC TGCCGGAGCC GTTGCGTGTG GCCAAGCCGA
4951 ATAAGTTTCA GTTTCTAGTG ATGGACTTGC CCATGGTGAT GGGCGACCGC
5001 CTCCATTGCA TGGATGTTCT CTTTGCTTTC ACTACCAGGG TCCTCGGGGA
5051 CTCCAGCGGC TTGGATACCA TGAAAACCAT GATGGAGGAG AAGTTTATGG
5101 AGGCCAACCC TTTTAAGAAG CTCTACGAGC CCATAGTCAC CACCACCAAG
5151 AGGAAGGAGG AGGAGCAAGG CGCCGCCGTC ATCCAGAGGG CCTACCGGAA
5201 ACACATGGAG AAGATGGTCA AACTGAGGCT GAAGGACAGG TCAAGTTCAT
5251 CGCACCAGGT GTTTTGCAAT GGAGACTTGT CCAGCTTGGA TGTGGCCAAG
5301 GTCAAGGTTC ACAATGACTG AACCCTCATC TAGA

24/27

Figure 6



This invention relates generally to sodium channel proteins and more particularly to a novel nucleic acid sequence encoding for a mammalian α -subunit of a voltage-gated, preferably tetrodotoxin-resistant, nervous tissue sodium channel protein. This invention further relates to its production by recombinant technology.

5 The basic unit of information transmitted from one part of the nervous system to another is a single action potential or nerve impulse. The „transmission line“ for these impulses is the axon, or nerve fiber. The electrical excitability of the nerve membrane has been shown to depend on the membrane's voltage-sensitive ionic permeability system that allows it to use energy stored in ionic concentration gradients. Electrical activity of the nerve
10 is triggered by a depolarization of the membrane, which opens channels through the membrane that are highly selective for sodium ions, which are then driven inward by the electrochemical gradient. Of the many ionic channels, the voltage-gated or voltage-sensitive sodium channel is one of the most studied. It is a transmembrane protein that is essential for the generation of action potentials in excitable cells. An excellent review of sodium channels is presented in
15 Catterall, TINS 16(12), 500-506 (1993).

 The cDNAs for several Na^+ channels have been cloned and sequenced. Numa *et al.*, Annals of the New York Academy of Sciences 479, 338-355 (1986), describe cDNA from the electric organ of eel and two different ones from rat brain. Rogart, U.S. Patent No. 5,380,836, describes cDNA from rat cardiac tissue. See also Rogart *et al.*, Proc. Natl. Acad. Sci. 86,
20 8170-8174 (1989). The sequence of PN1 and its orthologs in humans (hNE) and rabbits (Na^+ s) have been published (see, for example, Klugbauer *et al.*, EMBOJ 14, 1084-1090 (1995) and Belcher *et al.*, Proc. Natl. Acad. Sci. U.S.A. 923, 11034-11038 (1995)). The sequence of rat PN1 cloned from DRG and its function expression have been described (see, for example, Sangameswaran *et al.*, J.Biol.Chem. 272, 14805-14809 (1997)). Other cloned sodium
25 channels include rat brain types I and II, Noda *et al.*, Nature 320, 188-192 (1986), IIa, Auld *et al.*, Neuron 1, 449-461 (1988), and III, Kayano *et al.*, FEBS Lett. 228, 187-194 (1988), rat

skeletal muscle (SkM1), Trimmer *et al.*, Neuron 3, 33-49 (1989), rat NaCh6, Schaller *et al.*, J. Neurosci. 15, 3231-3242 (1995), rat peripheral nerve sodium channel type 3 (rPN3), Sangameswaran *et al.*, J. Biol Chem. 271, 5953-5956 (1996), also called SNS, Akopian *et al.*, Nature 379, 257-262 (1996), rat atypical channel, Felipe *et al.*, J. Biol. Chem. 269, 30125-30131 (1994), and the rat glial sodium channel, Akopian *et al.*, FEBS Lett. 400, 183-187 (1997).

These studies have shown that the amino acid sequence of the Na⁺ channel has been conserved over a long evolutionary period. These studies have also revealed that the channel is a single polypeptide containing four internal repeats, or homologous domains (domains I-IV), having similar amino acid sequences. Each domain folds into six predicted and helical transmembrane segments: five are hydrophobic segments and one is highly charged with many positively charged lysine and arginine residues. This highly charged segment is the fourth transmembrane segment in each domain (the S4 segment) and is likely to be involved in voltage-gating. The positively charged side chains on the S4 segment are likely to be paired with the negatively charged side chains on the other five segments such that membrane depolarization could shift the position of one helix relative to the other, thereby opening the channel. Accessory subunits may modify the function of the channel.

Therapeutic utility in recombinant materials derived from the DNA of the numerous sodium channels have been discovered. For example, U.S. Patent No. 5,132,296 by Cherksey discloses purified Na⁺ channels that have proven useful as therapeutic and diagnostic tools.

Isoforms of sodium channels are divided into „subfamilies“. The term „isoform“ is used to mean distinct but closely related sodium channel proteins, i.e., those having an amino acid homology of approximately 60-80%. These also show strong homology in functions. The term „subfamilies“ is used to mean distinct sodium channels that have an amino acid homology of approximately 80-95%. Combinations of several factors are used to determine the distinctions within a subfamily, for example, the speed of a channel, chromosomal location, expression data, homology to other channels within a species, and homology to a

are strongly expressed in adult DRG and nodose ganglia, less strongly expressed in brain, spinal cord and superior cervical ganglia, and not expressed in sciatic nerve, heart or skeletal muscle. In presently preferred forms, novel DNA sequences comprise cDNA sequences encoding rat nervous tissue sodium channel protein. One aspect of the present invention is the
5 α -subunit of this sodium channel protein.

Disclosed is the DNA, cDNA, and mRNA derived from the nucleic acid sequences of the invention and the cRNA derived from the mRNA. Specifically, two cDNA sequences together encode for the full length rat nervous tissue sodium channel.

Also included in this invention are alternate DNA forms, such as genomic DNA, DNA
10 prepared by partial or total chemical synthesis from nucleotides, and DNA having deletions or mutations.

Still another aspect of the invention is the novel rat TTX-resistant sodium channel protein and fragments thereof, encoded by the DNA of this invention.

Another aspect of the present invention are recombinant polynucleotides and
15 oligonucleotides comprising a nucleic acid sequence derived from the DNA sequence of this invention.

Another aspect of the invention is a method of stabilizing the full length cDNA which encodes the protein sequence of the invention.

Further aspects of the invention include expression vectors comprising the DNA of the
20 invention, host cells transformed or transfected by these vectors, and a cDNA library of these host cells.

Also forming part of this invention is an assay for inhibitors of the sodium channel protein comprising contacting a compound suspected of being an inhibitor with expressed sodium channel and measuring the activity of the sodium channel.

25 Further provided is a method of inhibiting the activity of the TTX-resistant sodium channel comprising administering an effective amount of a compound having an IC_{50} of 10 μ M or less.

Additionally provided are methods of employing the DNA for forming monoclonal and polyclonal antibodies, for use as molecular targets for drug discovery, highly specific markers for specific antigens, detector molecules, diagnostic assays, and therapeutic uses, such as pain relief, a probe for the PN5 channel in other mammalian tissue, designing therapeutics and screening for therapies.

BRIEF DESCRIPTION OF THE SEQ ID'S AND FIGURES

Figures 1A-E depict the 5908 nucleotide cDNA native sequence encoding the rat sodium channel type 5 („PN5“) (SEQ ID NO: 1), derived from two overlapping cDNA clones, designated 26.2 and 1.18.

Figures 2A-F depict the deduced amino acid sequence of PN5 (SEQ ID NO: 2, represented in the three-letter amino acid code). Figures 2G-H, depicting the deduced amino acid sequence of PN5 in single letter amino acid code, also show the homologous domains (I-IV); the putative transmembrane segments (S1-S6); the amino acid conferring resistance to TTX (♦); N-glycosylation sites (•); cAMP-dependent protein kinase A (PKA) phosphorylation site (O); and the termination codon (*).

Figure 3A depicts an 856 base pair sequence for the human PN5 (SEQ ID NO: 3).

Figure 3B depicts the amino acid sequence comparison of the hPN5 fragment with rat PN5.

Figure 4 depicts the sequence for the novel sodium channel domain IV probe (SEQ ID NO: 4).

Figures 5A-E depict the 5334 nucleotide sequence modified for stability and expression (SEQ ID NO: 5). Nucleotides 24 to 5518 constitute the 5295 bp region coding for a 1765 amino acid protein.

Figure 6 depicts the cloning map of PN5.

DETAILED DESCRIPTION OF THE INVENTION

The present invention relates to a purified and isolated nucleic acid sequence encoding for a novel mammalian, preferably TTX-resistant, sodium channel protein. The term "purified

and isolated DNA" refers to DNA that is essentially free, i.e. contains less than about 30%, preferably less than about 10%, and even more preferably less than about 1%, of the DNA with which the DNA of interest is naturally associated. Techniques for assessing purity are well known to the art and include, for example, restriction mapping, agarose gel

5 electrophoresis, and CsCl gradient centrifugation.

The term "DNA" is meant to include „cDNA“, or complementary DNA, which is single-stranded or double-stranded DNA sequences made by reverse transcription of mRNA isolated from a donor cell or by chemical synthesis. For example, treatment of mRNA with a reverse transcriptase such as AMV reverse transcriptase or M-MuLV reverse transcriptase in
10 the presence of an oligonucleotide primer will furnish an RNA-DNA duplex which can be treated with RNase H, DNA polymerase, and DNA ligase to generate double-stranded cDNA. If desired, the double-stranded cDNA can be denatured by conventional techniques such as heating to generate single-stranded cDNA. The term „cDNA“ includes cDNA that is a complementary copy of the naturally occurring mRNA ,as well as complementary copies of
15 variants of the naturally occurring mRNA that have the same biological activity. Variants would include, for example, insertions, deletions, sequences with degenerate codons and alleles.

„cRNA“ corresponding to mRNA transcribed from a DNA sequence encoding the α -subunit of a novel, preferably TTX-resistant, sodium channel protein is contemplated by this
20 invention. The term „cRNA“ refers to RNA that is a copy of the mRNA transcribed by a cell.

Specifically, the invention encompasses DNA having the native versions of the nucleotide sequences set forth in Figures 1A-E (SEQ ID NO: 1) designated herein as sodium channel type 5 (PN5). Figures 1A-E depict the 5908 nucleotide cDNA construct comprising a 5298-base (counting the stop codon) open reading frame (SEQ ID NO:1). Nucleotide residue
25 79 represents the start site of translation and residue 5376 represents the end of the stop codon.

The invention also encompasses engineered versions of PN5, and specifically the version as set forth in Figures 5A-E (SEQ ID NO: 5). This 5334 nucleotide SaII-XbaI clone

lacks most of the untranslated sequences, the 5298 nucleotide open reading frame beginning at nucleotide 24 and ending at nucleotide 5321. The start and stop codons are underlined, as are the translationally silent mutations at nucleotides 3932, 3935, 3941, 3944, and 3947, which were introduced to block rearrangement in this region during growth in *E. Coli*.

5 The nucleotide sequence of SEQ ID NO: 1 (Figures 1A-E) corresponds to the cDNAs from rat. A homology search provided that the closest related sodium channel is found in the rat cardiac channel, with 72.5% homology. The next closely related channels are rPN1, with 72% and rat brain types I and III, with 71.8% and 71.3% respectively. Homology to rPN3a, hPN3, rPN4, rPN4a, rat brain type II and rat skeletal muscle are each approximately 70 to
10 71%.

 Additionally, an 856 base pair clone (SEQ ID NO: 3) as shown in Figure 3A has been isolated from a human dorsal root ganglia (DRG) „cDNA library“ and is closely related to the rat PN5 amino acid sequence with 79% identity and 86% homology. The human PN5 sequence spans the region between IIIS1 and interdomain III/IV which includes the fast
15 inactivation gate (i.e., IFM) that is located within interdomain III/IV.

 The term „cDNA library“ refers to a collection of clones, usually in a bacteriophage, or less commonly in bacterial plasmids, containing cDNA copies of mRNA sequences derived from a donor cell or tissue.

 It is believed that additional homologs of the novel rat TTX-resistant sodium channel
20 described herein are also expressed in other mammalian tissue.

 Northern blot analysis (Example 5) indicates that PN5 is encoded by a ~6.5 kb transcript.

 The deduced amino acid sequence of PN5, shown in Figures 2A-F (SEQ ID NO: 2), exhibits the primary structural features of an α -subunit of a voltage-gated, TTX-resistant
25 sodium channel. Shown in Figures 2G-H are the homologous domains (I-IV); the putative transmembrane segments (S1-S6); the amino acid conferring resistance to TTX (\blacklozenge); N-glycosylation sites (\bullet); and cAMP-dependent PKA phosphorylation sites (O). DNA sequences

encoding the same or allelic variant or analog sodium channel protein polypeptides of the nervous system, through use of, at least in part, degenerate codons are also contemplated by this invention.

An interesting feature of this deduced amino acid sequence is that the amino acid that is most responsible for TTX-sensitivity is located at position 355 and is not aromatic. In rat and human brain type sodium channels, skeletal muscle channel, and in PN1 and PN4, this amino acid is tyrosine or phenylalanine and these channels are all TTX-sensitive. In PN3 and PN5, the amino acid is a serine. Since PN3 is highly resistant to TTX, the implication is that PN5 is also a TTX-resistant channel. The cardiac channel has a cysteine at this position and is „insensitive“ to TTX.

Although PN5 contains all of the hallmark features of a voltage-gated sodium channel, it has unique structural features that distinguish it from other sodium channels. For example, DIIS4 has 5 basic amino acids conserved in all sodium channels that could play a significant role in the voltage sensing aspects of the channel function. In PN5, the first basic amino acid is replaced by an alanine. Similarly, in DIIS4, PN5 has 5 basic amino acids rather than six that are present in other sodium channel sequences, the last arginine replaced by a glutamine. In DIIS3, the transmembrane segment contains only 18 amino acids, in contrast to 22 amino acids in other channels. Also, the short linker (4 amino acids) loop between S3 and S4 in DIIS is even shorter by a „deletion“ of 3 amino acids. This shortening of the S3 and the linker loop has been confirmed by designing primers in the appropriate region of the sequence for an RT-PCR experiment from rat DRG and sequencing the amplified DNA fragment. Such an experiment has been performed to confirm the sequence of another region of PN5, in the DIVS5-S6 loop, where there was a deletion of an 8 amino acid peptide.

Reverse transcription-polymerase chain reaction (oligonucleotide-primed RT-PCR) tissue distribution analysis of RNA from the rat central and peripheral nervous systems, in particular from rat DRG, was performed. Eight main tissue types were screened for expression of the unique PN5 genes corresponding to positions 5651-5903 of SEQ ID NO: 1

(Figures 1A-E). PN5 mRNA was present in five of the tissues studied: brain, spinal cord, DRG, nodose ganglia, and superior cervical ganglia. PN5 was not present in the remaining tissues studied: sciatic nerve tissue, heart or skeletal muscle tissue. PN5 was found to be the strongest in DRG and nodose ganglia, leading the applicants to believe that the DRG is enriched with PN5. PN5 shows dramatic abundance differences across a range of tissues. PN5 has a gradient of expression with high expression in DRG. PN5 has a gradient of expression like other channels, but more limited distribution.

The invention not only includes the entire protein expressed by the cDNA sequences of SEQ ID NOS: 1, 2 and 3, but also includes protein fragments. These fragments can be obtained by cleaving the full length proteins or by using smaller DNA sequences or „polynucleotides“ to express the desired fragment.

The term "polynucleotide" as used herein refers to a polymeric form of nucleotides of any length, either ribonucleotides or deoxyribonucleotides. This term refers only to the primary structure of the molecule. Thus, this term includes double- and single-stranded DNA, as well as double- and single-stranded RNA. It also includes modified, for example, by methylation and/or by capping, and unmodified forms of the polynucleotide.

Further, the term "polynucleotide" is intended to include a recombinant polynucleotide, which is of genomic, cDNA, semisynthetic or synthetic origin which, by virtue of its origin or manipulation is not associated with all or a portion of the polynucleotide with which it is associated in nature and/or is linked to a polynucleotide other than that to which it is linked in nature.

Accordingly, the invention also includes polynucleotides that can be used to make polypeptides of about 10 to 1500, preferably 10 to 100, amino acids in length. The isolation and purification of such recombinant polypeptides can be accomplished by techniques that are well known in the art, for example, preparative chromatographic separations or affinity chromatography. In addition, polypeptides can also be made by synthetic means which are well known in the art.

The invention allows for the manipulation of genetic materials by recombinant technology to produce polypeptides that possess the structural and functional characteristics of the novel voltage-gated, TTX-resistant sodium channel α -subunit found in sensory nerves.

Site directed mutagenesis can be used to provide such recombinant polypeptides. For example, synthetic oligonucleotides can be specifically inserted or substituted into the portion of the gene of interest to produce genes encoding for and expressing a specific mutant.

Random degenerate oligonucleotides can also be inserted and phage display techniques can be used to identify and isolate polypeptides possessing a functional property of interest.

In addition, the present invention contemplates recombinant polynucleotides of about 15 to 20kb, preferably 10 to 15kb, nucleotides in length, comprising a nucleic acid sequence „derived from“ the DNA of the invention.

The term "derived from" a designated sequence, refers to a nucleic acid sequence that is comprised of a sequence of approximately at least 6 to 8 nucleotides, more preferably at least 10 to 12 nucleotides, and, even more preferably, at least 15 to 20 nucleotides that correspond to, i.e., are homologous or complementary to, a region of the designated sequence. The derived sequence is not necessarily physically derived from the nucleotide sequence shown, but may be derived in any manner, including for example, chemical synthesis or DNA replication or reverse transcription, which are based on the information provided by the sequences of bases in the region(s) from which the polynucleotide is derived.

A neonatal expression test was performed with F11, a fusion cell line designed from neonatal rat DRG fused with a mouse cell line, N18TG, from Massachusetts General Hospital. F11 responds to trophic agents, such as NGF, by extending dendrites. It was found that PN5 was present in both native F11 and F11 treated with NGF, leading the applicants to believe that the sodium channel is natively expressed in F11.

In situ hybridization of PN5 mRNA to rat DRG tissue provides localization predominantly in the small and medium neurons with no detection in large neurons.

PN5 was also mapped to its cytogenetic location on mouse chromosome preparations. PN5 maps to the same chromosome as the cardiac channel and PN3.

In general, sodium channels comprise an α - and two β -subunits. The β -subunits may modulate the function of the channel. However, since the α -subunit is all that is required for the channel to be fully functional, expression of the cDNA in SEQ ID NO: 1 (Figures 1A-E) will provide a fully functional protein. The gene encoding the β_1 -subunit in peripheral nerve tissue was found to be identical to that found in rat heart, brain and skeletal muscle. The cDNA of the β_1 -subunit is not described herein as it is well known in the art, see Isom *et al.*, Neuron 12, 1183-1194 (1994). However, it is to be understood that by combining the known sequence for the β_1 -subunit with the α -subunit sequence described herein, one may obtain complete PN5 voltage-gated, preferably TTX-resistant, sodium channel.

The present invention also includes „expression vectors“ comprising the DNA or the cDNA described above, host cells transformed with these expression vectors capable of producing the sodium channel of the invention, and cDNA libraries comprising such host cells.

The term "expression vector" refers to any genetic element, e.g., a plasmid, a chromosome, a virus, behaving either as an autonomous unit of polynucleotide expression within a cell or being rendered capable of replication by insertion into a host cell chromosome, having attached to it another polynucleotide segment, so as to bring about the replication and/or expression of the attached segment. Suitable vectors include, but are not limited to, plasmids, bacteriophages, and cosmids. Vectors will contain polynucleotide sequences which are necessary to effect ligation or insertion of the vector into a desired host cell and to effect the expression of the attached segment. Such sequences differ depending on the host organism, and will include promoter sequences to effect transcription, enhancer sequences to increase transcription, ribosomal binding site sequences and transcription and translation termination sequences.

The term "host cell" generally refers to prokaryotic or eukaryotic organisms and includes any transformable or transfectable organism which is capable of expressing a protein and can be, or has been, used as a recipient for expression vectors or other transferred DNA.

Host cells can also be made to express protein by direct injection with exogenous cRNA
5 translatable into the protein of interest. A preferred host cell is the *Xenopus* oocyte.

The term "transformed" refers to any known method for the insertion of foreign DNA or RNA sequences into a host prokaryotic cell. The term „transfected" refers to any known method for the insertion of foreign DNA or RNA sequences into a host eukaryotic cell. Such transformed or transfected cells include stably transformed or transfected cells in which the
10 inserted DNA is rendered capable of replication in the host cell. They also include transiently expressing cells which express the inserted DNA or RNA for limited periods of time. The transformation or transfection procedure depends on the host cell being transformed. It can include packaging the polynucleotide in a virus as well as direct uptake of the polynucleotide, such as, for example, lipofection or microinjection. Transformation and transfection can result
15 in incorporation of the inserted DNA into the genome of the host cell or the maintenance of the inserted DNA within the host cell in plasmid form. Methods of transformation are well known in the art and include, but are not limited to, viral infection, electroporation, lipofection, and calcium phosphate mediated direct uptake.

It is to be understood that this invention is intended to include other forms of
20 expression vectors, host cells, and transformation techniques which serve equivalent functions and which become known to the art hereto.

The invention also pertains to an assay for inhibitors of the novel TTX-resistant sodium channel protein comprising contacting a compound suspected of being an inhibitor with expressed sodium channel and measuring the activity of the sodium channel. The
25 compound can be a substantially pure compound of synthetic origin combined in an aqueous medium, or the compound can be a naturally occurring material such that the assay medium is an extract of biological origin, such as, for example, a plant, animal, or microbial cell extract.

PN5 activity can be measured by methods such as electrophysiology (two electrode voltage clamp or single electrode whole cell patch clamp), guanidinium ion flux assays, and toxin-binding assays. An "inhibitor" is defined as generally that amount that results in greater than 50% decrease in PN5 activity, preferably greater than 70% decrease in PN5 activity, more preferably greater than 90% decrease in PN5 activity.

Many uses of the invention exist, a few of which are described below:

1. Probe for mamalian channels.

As mentioned above, it is believed that additional homologs of the novel rat TTX-resistant sodium channel described herein are also expressed in mammalian tissue, in particular, human tissue. The entire cDNAs of PN5 rat sodium channels of the present invention can be used as a probe to discover whether additional novel PN5 voltage-gated, preferably TTX-resistant, sodium channels exist in human tissue and, if they do, to aid in isolating the cDNAs for the human protein.

The human homologues of the rat TTX-resistant PN5 channels can be cloned using a human DRG cDNA library. Human DRG are obtained at autopsy. The frozen tissue is homogenized and the RNA extracted with guanidine isothiocyanate (Chirgwin *et al.* Biochemistry 18, 5294-5299, (1979)). The RNA is size-fractionated on a sucrose gradient to enrich for large mRNAs because the sodium channel α -subunits are encoded by large (7-11 kb) transcripts. Double-stranded cDNA is prepared using the SuperScript Choice cDNA kit (GIBCO BRL) with either oligo(dT) or random hexamer primers. EcoRI adapters are ligated onto the double-stranded cDNA which is then phosphorylated. The cDNA library is constructed by ligating the double-stranded cDNA into the bacteriophage-lambda ZAP II vector (Stratagene) followed by packaging into phage particles.

Phage are plated out on 150 mm plates on a lawn of XLI-Blue MRF' bacteria (Stratagene) and plaque replicas are made on Hybond N nylon membranes (Amersham). Filters are hybridized to rat PN5 cDNA probes by standard procedures and detected by autoradiography or chemiluminescence. The signal produced by the rat PN5 probes

hybridizing to positive human clones at high stringency should be stronger than obtained with rat brain sodium channel probes hybridizing to these clones. Positive plaques are further purified by limiting dilution and re-screened by hybridization or PCR. Restriction mapping and polymerase chain reaction will identify overlapping clones that can be assembled by
5 standard techniques into the full-length human homologue of rat PN5. The human clone can be expressed by injecting cRNA transcribed *in vitro* from the full-length cDNA clone into *Xenopus* oocytes, or by transfecting a mammalian cell line with a vector containing the cDNA linked to a suitable promoter.

2. Antibodies Against PN5.

10 The polypeptides of the invention are highly useful for the development of antibodies against PN5. Such antibodies can be used in affinity chromatography to purify recombinant sodium channel proteins or polypeptides, or they can be used as a research tool. For example, antibodies bound to a reporter molecule can be used in histochemical staining techniques to identify other tissues and cell types where PN5 are present, or they can be used to identify
15 epitopic or functional regions of the sodium channel protein of the invention.

The antibodies can be monoclonal or polyclonal and can be prepared by techniques that are well known in the art. Polyclonal antibodies are prepared as follows: an immunogenic conjugate comprising PN5 or a fragment thereof, optionally linked to a carrier protein, is used to immunize a selected mammal such as a mouse, rabbit, goat, etc. Serum from the
20 immunized mammal is collected and treated according to known procedures to separate the immunoglobulin fraction.

Monoclonal antibodies are prepared by standard hybridoma cell technology based on that reported by Kohler and Milstein in Nature 256, 495-497 (1975). Spleen cells are obtained from a host animal immunized with the PN5 protein or a fragment thereof, optionally linked to
25 a carrier. Hybrid cells are formed by fusing these spleen cells with an appropriate myeloma cell line and cultured. The antibodies produced by the hybrid cells are screened for their ability to bind to expressed PN5 proteins.

A number of screening techniques well known in the art, such as, for example, forward or reverse enzyme-linked immunosorbent assay screening methods, may be employed. The hybrid cells producing such antibodies are then subjected to recloning and high dilution conditions in order to select a hybrid cell that secretes a homogeneous population of antibodies
5 specific to either the PN5 protein.

In addition, antibodies can be raised by cloning and expressing nucleotide sequences or mutagenized versions thereof coding at least for the amino acid sequences required for specific binding of natural antibodies, and these expressed proteins used as the immunogen. Antibodies may include the complete immunoglobulin or a fragment thereof. Antibodies may
10 be linked to a reporter group such as is described above with reference to polynucleotides.

Example 10 illustrates practice of producing an antibody.

3. Therapeutic Targets for Compounds to Treat Disorders and Assays Thereof.

The present invention also includes the use of the novel voltage-gated, preferably TTX-resistant, sodium channel α -subunit as a therapeutic target for compounds to treat disorders of
15 the nervous system based on the RT-PCR localization data. The disorders include, but are not limited to, epilepsy, stroke injury, brain injury, diabetic neuropathy, traumatic injury, chronic neuropathic pain, and AIDS-associated neuropathy.

4. Designing Therapeutics based on Inhibiting PN5 and assays thereof.

This invention is also directed to inhibiting the activity of PN5 in brain, spinal cord,
20 DRG, nodose ganglia, and superior cervical ganglia tissues. However, it is to be understood that further studies may reveal that PN5 is present in other tissues, and as such, those tissues can also be targeted areas. For example, the detection of PN5 mRNA in nodose ganglia suggests that PN5 may conduct TTX-resistant sodium currents in this and other sensory ganglia of the nervous system.

25 In addition, it has been found that proteins not normally expressed in certain tissues are expressed in a disease state. Therefore, this invention is intended to encompass the inhibition

of PN5 in tissues and cell types where the protein is normally expressed, and in those tissues and cell types where the protein is only expressed during a disease state.

For example, it is believed that TTX-resistant sodium channels play a key role in transmitting nerve impulses relating to sensory inputs such as pain and pressure. This information will facilitate the design of therapeutics that can be targeted to a specific area such as peripheral nerve tissue.

The recombinant protein of the present invention can be used to screen for potential therapeutics that have the ability to inhibit the sodium channel of interest. In particular, it would be useful to inhibit selectively the function of sodium channels in peripheral nerve tissues responsible for transmitting pain and pressure signals without simultaneously affecting the function of sodium channels in other tissues such as heart and muscle. Such selectivity would allow for the treatment of pain without causing side effects due to cardiac or neuromuscular complications. Therefore, it would be useful to have DNA sequences coding for sodium channels that are selectively expressed in peripheral nerve tissue.

5. Pain Reliever.

Sodium channels in peripheral nerve tissue play a large role in the transmission of nerve impulses, and therefore are instrumental in understanding neuropathic pain transmission. Neuropathic pain falls into two components: allodynia, where a normally non-painful stimulus becomes painful, and hyperalgesia, where a usually normal painful stimulus becomes extremely painful.

In tissue localization studies, PN5 mRNA maps small and medium neurons of DRG. PN5 mRNA is also present in brain and spinal cord. Inhibiting its activities may help prevent ailments such as headaches and migraines. The ability to inhibit the activity of these sodium channels, i.e., reduce the conduction of nerve impulses, will affect the nerve's ability to transmit pain impulses. Selective inhibition of sodium channels in sensory neurons such as DRG will allow the blockage of pain impulses without complicating side effects caused by inhibition of sodium channels in other tissues such as brain and heart. In addition, certain

diseases are caused by sodium channels that produce impulses at an extremely high frequency. The ability to reduce the activity of the channel can then eliminate or alleviate the disease. Accordingly, potential therapeutic compounds can be screened by methods well known in the art to discover whether they can inhibit the activity of the recombinant sodium channel of the invention. Barram, M. *et al.*, Naun-Schmiedeberg's Archives of Pharmacology 347, 125-132 (1993) and McNeal, E.T. *et al.*, J. Med. Chem. 28, 381-388 (1985). For similar studies with the acetyl choline receptor, see, Claudio *et al.*, Science 238, 1688-1694 (1987).

For example, pain can be alleviated by inhibiting the activity of the novel preferably TTX-resistant sodium channel comprising administering a therapeutically effective amount of a compound having an IC_{50} approximately 10 μM or less, preferably $\leq 1 \mu M$. Potential therapeutic compounds are identified based on their ability to inhibit the activity of PN5. Therefore, the aforementioned assay can be used to identify compounds having a therapeutically effective IC_{50} .

The term „ IC_{50} “ refers to the concentration of a compound that is required to inhibit by 50% the activity of expressed PN5 when activity is measured by electrophysiology, flux assays, and toxin-binding assays, as mentioned above.

6. Diagnostic Assays.

The basic molecular biology techniques employed in accomplishing features of this invention, such as RNA, DNA and plasmid isolation, restriction enzyme digestion, preparation and probing of a cDNA library, sequencing clones, constructing expression vectors, transforming cells, maintaining and growing cell cultures, and other general techniques are well known in the art, and descriptions of such techniques can be found in general laboratory manuals such as Molecular Cloning: A Laboratory Manual by Sambrook *et al.* (Cold Spring Harbor Laboratory Press, 2nd edition, 1989).

For example, the polynucleotides of the invention can be bound to a „reporter molecule“ to form a polynucleotide probe useful for Northern and Southern blot analysis and *in situ* hybridizations.

The term "reporter molecule" refers to a chemical entity capable of being detected by a suitable detection means, including, but not limited to, spectrophotometric, chemiluminescent, immunochemical, or radiochemical means. The polynucleotides of this invention can be conjugated to a reporter molecule by techniques well known in the art. Typically the reporter molecule contains a functional group suitable for attachment to or incorporation into the polynucleotide. The functional groups suitable for attaching the reporter group are usually activated esters or alkylating agents. Details of techniques for attaching reporter groups are well known in the art. See, for example, Matthews, J.A., Batki, A., Hynds, C., and Kricka, L.J., *Anal. Biochem.* 151, 205-209 (1985) and Engelhardt *et al.*, European Patent Application No. 0302175.

Accordingly, the following Examples are merely illustrative of the techniques by which the invention can be practiced.

Abbreviations

The following abbreviations are used throughout the Examples and have each of the respective meanings defined below.

BSA: bovine serum albumin

Denhardt's solution: 0.02% BSA, 0.02% polyvinyl-pyrrolidone, 0.02% Ficoll (0.1 g BSA, 0.1 g Ficoll and 0.1 g polyvinylpyrrolidone per 500 ml)

DRG: dorsal root ganglia

EDTA: Ethylenediaminetetraacetic acid, tetrasodium salt

MEN: 20 mM MOPS, 1 mM EDTA, 5 mM sodium acetate, pH 7.0

MOPS: 3-(N-morpholino)propanesulfonic acid (Sigma Chemical Company)

PN5: peripheral nerve sodium channel 5

PNS: peripheral nervous system

SDS: sodium dodecyl sulfate

SSC: 150 mM NaCl, 15 mM sodium citrate, pH 7.0

SSPE: 80 mM NaCl, 10 mM sodium phosphate, 1 mM ethylenediaminetetraacetate, pH :

8.0

TEV: two electrode voltage clamp

TTX: tetrodotoxin (Sigma Chemical Company)

EXAMPLES

The following Examples illustrate practice of the invention.

Materials

The plasmid pBK-CMV was obtained from Stratagene (La Jolla, CA); the plasmid
5 pBSTA is described by Goldin *et al.*, in Methods in Enzymology (Rudy & Iverson, eds.) 207,
279-297; the plasmid pCIneo was obtained from Promega (Madison, WI); and the plasmid
pCRII was obtained from Invitrogen (Carlsbad, CA).

The oocyte expression vector plasmid pBSTAcIIr was constructed from
pBSTA by insertion of a synthetic oligonucleotide linker; plasmid pKK232-8 was obtained
10 from Pharmacia Biotech (Piscataway, NJ); plasmid pCRII was obtained from Invitrogen, San
Diego, CA. Competent *E. coli* cell lines STBL2™ and SURE® were obtained from
Gibco/BRL and Stratagene, respectively.

EXAMPLE 1

OBTAINING RNA FROM RAT DRG, BRAIN AND SPINAL CORD

15

Lumbar DRG No. 4 and No. 5 (L4 and L5) brain and spinal cord were removed from
anesthetized adult male Sprague-Dawley rats under a dissecting microscope. The tissues were
frozen in dry ice and homogenized with a Polytron homogenizer; the RNA was extracted by
the guanidine isothiocyanate procedure (see Chomczynski *et al.*, Anal. Biochemistry 162: 156-
20 159 (1987)). Total RNA (5 µg of each sample) was dissolved in MEN buffer containing 50%
formamide, 6.6% formaldehyde and denatured at 65°C for 5-10 min. The RNA was
electrophoresed through a 0.8% agarose gel containing 8.3% formaldehyde in MEN buffer.
The electrode buffer was MEN buffer containing 3.7% formaldehyde; the gel was run at 50 V
for 12-18 hours.

25

Size markers, including ribosomal 18S and 28S RNAs and RNA markers (GIBCO
BRL), were run in parallel lanes of the gel. Their positions were determined by staining the
excised lane with ethidium bromide (0.5 µg/ml) followed by photography under UV light.

After electrophoresis, the gel was rinsed in 2xSSC and the RNA was transferred to a Duralose membrane (Stratagene) with 20xSSC by capillary action; the membrane was baked under vacuum at 80°C for 1 hour.

5

EXAMPLE 2

PROBE FROM RAT BRAIN IIA

A ³²P-labeled cRNA probe complementary to nucleotides 4637-5868 of the rat brain IIA sodium channel α -subunit sequence was synthesized *in vitro* with T7 RNA polymerase (Pharmacia) using pEAF8 template DNA, (Noda *et al.*, Nature 320, 188-192 (1986)) that had been linearized with BstEII.

Protocols for each procedure mentioned above can be found in Molecular Cloning: A Laboratory Manual by Sambrook *et al.* (Cold Spring Harbor Laboratory Press, 2nd edition, 1989).

15

EXAMPLE 3

HYBRIDIZATION OF RNA WITH THE PROBE FROM RAT BRAIN IIA

The membrane of Example 1 was prehybridized in 50% formamide, 5xSSC, 50 mM sodium phosphate, pH 7.1, 1x Denhardt's solution, 0.5% SDS, and sheared, heat-denatured salmon sperm DNA (1 mg/ml) for 16 hours at 42°C. The membrane was hybridized in 50% formamide, 5xSSC, 50 mM sodium phosphate, pH 7.1, 1x Denhardt's solution, 0.5% SDS, and sheared, heat-denatured salmon sperm DNA (200 μ g/ml) with the ³²P-labeled cRNA probe (ca. $1-3 \times 10^6$ cpm/ml) described in Example 2 for 18 hours at 42°C.

25

The membrane was rinsed with 2xSSC, 0.1% SDS at room temperature for 20 min. and then washed sequentially with: 2xSSC, 0.1% SDS at 55°C for 30 min., 0.2xSSC, 0.1% SDS at 65°C for 30 min., 0.2xSSC, 0.1% SDS at 70°C for 30 min., and 0.2xSSC, 0.1% SDS, 0.1% sodium pyrophosphate at 70°C for 20 min. The filter was exposed against Kodak X-omat AR film at -80°C with intensifying screens for up to 2 weeks.

The pEAF8 probe hybridized to mRNAs in the DRG sample with sizes of 11 kb, 9.5 kb, 7.3 kb, and 6.5 kb, estimated on the basis of their positions relative to the standards.

EXAMPLE 4

5 NOVEL SODIUM CHANNEL DOMAIN IV PROBE

The probe was obtained as follows: RT-PCR was performed on RNA isolated from rat DRG using degenerate oligonucleotide primers that were designed based on the homologies between known sodium channels in domain IV. The domain IV products were cloned into a
10 plasmid vector, transformed into *E. coli* and single colonies isolated. The domain IV specific PCR products obtained from several of these colonies were individually sequenced. Cloned novel domain IV sequence was as follows (SEQ ID NO: 4):

```

1      CTCAACATGG TTACGATGAT GGTGGAGACC GACGAGCAGG GCGAGGAGAA
51     GACGAAGGTT CTGGGCAGAA TCAACCAGTT CTTTGTGGCC GTCTTCACGG
15  101  GCGAGTGTGT GATGAAGATG TTCGCCCTGC GACAGTACTA TTTCACCAAC
151   GGCTGGAACG TGTTCGACTT CATAGTGGTG ATCCTGTCCA TTGGGAGTCT
201   GCTGTTTCT  GCAATCCTTA AGTCACTGGA AACTACTTC TCCCCGACGC
251   TCTTCCGGGT CATCCGTCTG GCCAGGATCG GCCGCATCCT CAGGCTGATC
301   CGAGCAGCCA AGGGGATTCG CACGCTGCTC TTCGCCCTCA TGATGTCCCT
20  351   GCCCCCCTC TTCAACATCG GCCTCCTCCT CTTCTCGTC ATGTTTCATCT
401   ACTCCATCTT CGGCATGGCC AGCTTCGCTA ACGTCGTGGA CGAGGCCGGC
451   ATCGACGACA TGTTCAACTT CAAGACCTTT GGCAACAGCA TGCTGTGCCT
501   GTTCCAGATC ACCACCTCGG CCGGCTGGGA CGGCCTCCTC AGCCCCATCC
551   TCAACACGGG GCCTCCCTAC TGCGACCCCA ACCTGCCCAA CAGCAACGGC
25  601   TCCCGGGGGA ACTGCGGGAG CCCGGCGGTG GGCATCATCT TCTTCACCAC
651   CTACATCATC ATCTCCTTCC TCATCGTGGT CAACATGTAT ATCGCAGTCA
701   TC

```

This sequence was labeled with ³²P by random priming.

30

EXAMPLE 5

HYBRIDIZATION OF RNA WITH THE NOVEL SODIUM CHANNEL 3'-UTR PROBE

5 A Northern blot was prepared with 10µg total RNA from rat brain, spinal cord, and
DRG. The blot was hybridized with a cRNA probe from the 3'-UTR. The 3'-UTR was
cloned into pSP 73 vector, the cRNA transcribed using a Trans Probe T kit (Pharmacia
Biotech) and ³²P UTP. The blot was prehybridized for 2 hours at 65°C in a solution
containing 5XSSC, 1X Denhardt's solution, 0.5% SDS, 50mM sodium phosphate, pH 7.1,
10 salmon sperm DNA (1mg/ml) and 50% formamide. Hybridization was conducted at 45°C for
18 hours in the above solution except that the salmon sperm DNA was included at a
concentration of 200µg/ml and the ³²P-labeled probe was added at 7.5x10⁵ cpm.ml solution.
The blot was subsequently washed three times at 2XSSC and 0.1% SDS at room temperature,
once with 0.2XSSC and 0.1% SDS at 65°C for 20 min., and once with 0.2XSSC, 0.1% SDS
15 and 0.1% sodium pyrophosphate at 65°C for 20 min. The blot was analyzed on a
PhosphoImager (BioRad) after an exposure of 2 days. The results indicated that there was a
~6.5kb band signal present in brain only in the lane containing RNA from DRG. Because of
the lower abundance of PN5 mRNA, as evidenced by the RT-PCR experiment, the 6.5kb band
was not detectable in brain and spinal cord.

20

EXAMPLE 6

CONSTRUCTION & SCREENING OF cDNA LIBRARY FROM RAT DRG

25 An EcoRI-adapted cDNA library was prepared from normal adult male Sprague-
Dawley rat DRG poly(A)+ RNA using the SuperScript Choice System (GIBCO BRL). cDNA
(>4 kb) was selected by sucrose gradient fractionation as described by Kieffer, Gene 109, 115-
119 (1991). The cDNA was then ligated into the Zap Express vector (Stratagene), and
packaged with the Gigapack II XL lambda packaging extract (Stratagene). Similarly, a >2kb
30 DRG cDNA library was synthesized.



Phage (3.5×10^5) were screened by filter hybridization with a ^{32}P -labeled probe (rBIIa, bases 4637-5868 as follows of Auld *et al.*, Neuron 1, 449-461 (1988)). Filters were hybridized in 50% formamide, 5X SSPE, 5X Denhardt's solution, 0.5% SDS, 250 $\mu\text{g/ml}$ sheared, denatured salmon sperm DNA, and 50 mM sodium phosphate at 42°C and washed in 0.5X SSC/0.1% SDS at 50°C.

Southern blots of EcoRI-digested plasmids were hybridized with the ^{32}P -labeled DNA probe, (SEQ ID NO: 4). The filters were then hybridized in 50% formamide, 6X SSC, 5X Denhardt's solution, 0.5% SDS, and 100 $\mu\text{g/ml}$ sheared, denatured salmon sperm DNA at 42°C and were washed in 0.1X SSC/0.1% SDS at 65°C.

Positive clones were excised in vivo into pBK-CMV using the ExAssist/XLOR system (Stratagene).

EXAMPLE 7

CLONES AND NUCLEOTIDE ANALYSIS

cDNA clones, 26.2 and 25.1 were isolated from the >4kb DRG cDNA library and clone 1.18 was isolated from the >2kb DRG cDNA library. By sequence analysis, 26.2 appeared to be a full-length cDNA encoding a novel sodium channel and 25.1 extended from domain II to the 3'-UTR. However, each had a deletion which truncated the coding region. Clone 1.18 had the 3'- untranslated region, in addition to the C-terminus of the deduced amino acid sequence of PN5. The construct in the expression vector, pBSTACIIr, consisted of sequences from 26.2 and 1.18.

PN5 homology to other known sodium channels was obtained using the GAP/Best Fit (GCG) program:

	Channel	% Similarity	% Identity
30	PN3a	71	54
	hPN3	71	55
	PN4	71	53
	PN4a	71	53



	PN1	72	55
	rat brain type I	72	55
	rat brain type II	71	54
	rat brain type III	71	54
5	rat cardiac channel	73	56
	rat skeletal muscle channel	71	53

Stabilizing the PN5 full length cDNA

10 A. Media, *E. coli* cell lines, and growth conditions:

Growth of fragments of PN5 could be accomplished under standard conditions; however growth of plasmids containing full length constructs of PN5 (in pCIneo, pBSTAcIIr, and other vectors) could not be accomplished without use of special growth media, conditions, and *E. coli* strains. The following proved to be optimal: (1) use of *E. coli* STBL2™ for
15 primary transformation following ligation reactions and for large scale culturing; (2) solid media was 1/2x FM (see below) plus 1x LB (Tryptone, 1%, Yeast Extract, 0.5%, NaCl, 0.5%), plus 15g/L agar, or 1x FM plus 1/2x LB; (3) liquid media optimally was 1x FM plus 1/2x LB; (4) carbenicillin, 100µg/ml, was used for all media, as it is metabolized less rapidly than ampicillin; (5) temperature for growth should be no greater than 30°C, usually 24-26°C; this
20 necessitated longer growth periods than normally employed, from 24 to 72 hours.

2x Freezing Medium (2xFM):

	K2HP04	12.6g
	Na3Citrate	0.9g
	MgSO4.7H2O	0.18g
25	(NH4)2SO4	1.8g
	KH2PO4	3.6g
	Glycerol	88g
	H2O	qs to 1L

2x FM and the remaining media components are prepared separately, sterilized by autoclaving,
30 cooled to at least 60°C, and added together to form the final medium. Carbenicillin is prepared

at 25mg/ml H₂O and sterilized by filtration. 2x FM was first described for preparation of frozen stocks of bacterial cells (Practical Methods in Molecular Biology, Schleif, R.F. and Wensink, P.C., Springer-Verlag, New York (1981) pp. 201-202).

5 B. Expression Vectors

In order to provide for increased stability of the full length cDNA, the oocyte expression vector pBSTAcIIr was modified to reduce plasmid copy number when grown in *E. coli* and to reduce possible read-through transcription from vector sequences that might result in toxic cryptic expression of PN5 protein, Brosius J., Gene 27, 151-160(1984). pBSTAcIIr
10 was digested with PvuII. The 755 bp fragment containing the T7 promoter, β -globin 5'UTR, the multiple cloning site, β -globin 3'UTR, and T3 promoter was ligated to the 3.6 kb fragment containing the replication origin, ampicillin resistance gene, *rrnBT*₁ and *rrnBT*₁T₂ transcription terminators from pKK232-8, which had been fully digested with SmaI and partially digested with PvuII and treated with shrimp intestinal phosphatase to prevent self
15 ligation. The resulting plasmid in which the orientation of the pBSTA fragment is such that the T7 promoter is proximal to the *rrnBT*₁ terminator was identified by restriction mapping and named pHQ8. As is the case with pBSTA, the direction of transcription of the ampicillin resistance gene and replication origin of pHQ8 is opposite to that of the gene expression cassette, and the presence of the *rrnB* T₁ terminator should reduce any remaining read-through
20 from the vector into the T7 promoter driven expression cassette.

C. Assembly of full length cDNA for expression

Since pBK-CMV.26.2 had a 58 bp deletion (corresponding to bp 4346 to 4403 of SEQ ID NO: 1) and the sequence of pBK-CMV.1.18 begins at bp 4180 of SEQ ID NO: 1, pBK-CMV.1.18 could be used to „repair“ pBK-CMV.26.2. A strategy was developed to assemble a
25 full length cDNA from clones pBK-CMV.26.2 and pBK-CMV.1.18 in three sections, truncating the 5' and 3' UTRs and introducing unique restriction sites at the 5' and 3' ends in the process. The 5' end

was generated by PCR from 26.2, truncating the 5' UTR by incorporating a SalI site just upstream of the start codon. The central section was a restriction fragment from 26.2. The 3' end was prepared by overlap PCR from both 26.2 and 1.18 and incorporating an XbaI site just downstream of the stop codon. These sections were digested at unique restriction sites and assembled in pBSTAcIIr. Although this construct appeared to have a correct sequence, upon recloning as a SalI to XbaI fragment into pCIneo, two types of isolates were found, one with a deletion and one with an 8 bp insertion. Reexamination of the pBSTAcIIr clone showed the sequence was „mixed“ in this region, so that the clone must have rearranged. The 8 bp insertion was found as a repeat of one of the members of an 8 bp duplication in the native sequence, forming a triple 8 bp repeat in the rearranged isolate. Numerous cloning attempts inevitably gave rise to this rearrangement. Overlap PCR was used to introduce silent mutations into one of the 8 bp repeats, and a fragment containing this region was included when the PN5 coding region was assembled into HQ8, the low-copy number version of pBSTAcIIr, to give plasmid HR-1. This sequence proved to be stable (see Figures 5A-E, SEQ ID NO: 5).

The 5' end fragment was prepared by PCR using pBK-CMV.26.2 DNA as template and primers 4999 (CTTGGTCTGACTCTAGATCAGGGTGAAGATGGAGGAG; SalI site underlined, PN5 homology in italics, corresponding to bp 58-77 of SEQ ID NO: 1, initiation codon in bold) and 4927 (GGGTTCAATGTGGTTTTATCT, corresponding to bp 1067 to 1047 of SEQ ID NO: 1), followed by gel purification, digestion with SalI and KpnI (KpnI site at pb 1003-1008, SEQ ID NO: 1), and gel purification.

The central 3.1 kb fragment was prepared by digestion of pBK-CMV.26.2 DNA with KpnI and AatII (AatII site at 4133-4138), followed by gel purification.

The 3' end fragment was prepared as follows: PCR using primers 4837 (TCTGGGAAGTTTGGAAG, corresponding to bp 3613 to 3629 of SEQ ID NO: 1) and 4931

(GACCACGAAGGCTATGTTGAGG, corresponding to bp 4239 to 4218 of SEQ ID NO: 1) on pBK-CMV.26.2 DNA as template gave a fragment of 0.6 kb. PCR using primers 4930 (CCTCAACATAGCCTTCGTGGTC, corresponding to bp 4218 to 4239 of SEQ ID NO: 1) and 4929 (GTCTTCTAGATGAGGGTTCAGTCATTGTG, XbaI site underlined, PN5 homology in italics, corresponding to pb 5386 to 5365 of SEQ ID NO: 1, stop codon in bold) on pBK-CMV.1.18 DNA as template gave a fragment of 1.2 kb, introducing a XbaI site 7 bp from the stop codon. Thus the 3' end of the 4837-4931 fragment exactly complements the 5' end of the 4930-4929 fragment. These two fragments were gel purified and a fraction of each combined as template in a PCR reaction using primers 4928 (CAAGCCTTTGTGTTTCGAC, corresponding to bp 4084 to 4101 of SEQ ID NO: 1) and 4929, to give a fragment of 1.3 kb. This fragment was gel purified, digested with AatII and XbaI, and the 1.2 kb fragment gel purified.

The 3' end fragment was cloned into AatII and XbaI digested pBSTAcIIr. One isolate was digested with Sall and KpnI and ligated to the 5' end fragment. The resulting plasmid, after sequence verification, was digested with KpnI and AatII and ligated to the central 3.1 kb fragment, to form pBSTAcIIr.PN5(clone 21). pBSTAcIIr.PN5 (clone 21) was digested with Sall and XbaI to release the 5.3 kb PN5 fragment which was cloned into Sall and XbaI digested pCIneoII. Multiple isolates were found, of which GPII-1, which was completely sequenced, was typical and contained an 8 bp insert. This CAGAAGAA, after pb 3994 of SEQ ID NO: 1, converted the direct repeat of this sequence at this location into a triple direct repeat, causing a shift in the reading frame. In an attempt to repair this defect, pBSTAcIIr.PN5 (clone 21) was digested with NheI (bp 2538-2543 SEQ ID NO: 1) and XhoI (bp 4828-4833, SEQ ID NO: 1) to give a 6.2 kb fragment and with AatII and XhoI to give a 0.7 kb fragment which were ligated to the 1.6 kb fragment resulting from digestion of pBK-CMV.26.2 with AatII and NheI. Although no isolates were found which were completely correct, one isolate, HA-4, had only a single base

change, deletion of the C at bp 4827 (SEQ ID NO: 1) adjacent to the XhoI site.

In order to prevent the 8 bp insertion rearrangement from occurring, three silent mutations were introduced in the 5' repeat, and two additional mutations in a string of Ts would also be introduced, as shown below (bp 3982 to 4014, SEQ ID NO: 1; mutation sites

5 underlined, 8 bp repeats in native sequence in italics):

native	GAC	ATT	TTT	ATG	ACA	GAA	GAA	CAG	AAG	AAA	TAT
	Asp	Ile	Phe	Met	Thr	Glu	Glu	Gln	Lys	Lys	Tyr
mutant	GAC	ATC	TTC	ATG	ACT	GAG	GAG	CAG	AAG	AAA	TAT

- 10 As isolate HA-4 had the native direct repeat sequence (as opposed to e.g. pBSTAcIIr.PN5 (clone 21)) and the region near the XhoI site defect would not be involved, it was used as template DNA for the following PCR reactions. Primer P5-3716S (CCGAAGCCAATGTAAACATTAGTAATTACTCGTG, corresponding to pb 3684 to 3716, SEQ ID NO: 1) was paired with primer P5-3969AS
- 15 (GCTCCTCAGTCATGAAGATGTCTTGGCCACCTAAC, correspond to bp 4003 to 3969, SEQ ID NO: 1, mutated bases are underlined) to give a 320 bp product. Primer P5-4017S (GGCCAAGACATCTTCATGACTGAGGAGCAGAAGAAATATTAC, corresponding to bp 3976 to 4017, SEQ ID NO: 1; mutated bases are underlined) was paired with primer P5-4247AS (CTCAAAGCAAAGACTTTGATGAGACACTCTATGG, corresponding to bp 4280
- 20 to 4247, SEQ ID NO: 1) to give a 305 bp product. The 3' end of the 320 bp fragment thus has a 28 bp exact match to the 5' end of the 305 bp fragment. The two bands were gel purified and a fraction of each combined in a new PCR reaction with primers P5-3716S and P5-4247AS to give a 597 bp product, which was T/A cloned into vector pCRII. Isolate HO-7 was found to have the desired sequence. A four-way ligation was performed to assemble the full-
- 25 length, modified PN5:

the oocyte expression vector HQ-8 was digested with SalI and XbaI to give a 4.4 kb vector fragment; GPII-1 was digested with SalI and MluI to give a 3.8 kb fragment containing the 5' half of PN5; HO-7 was digested with MluI (bp 3866 to 3871, SEQ ID NO: 1) and AatII to give a 0.3 kb fragment containing the mutant 8 bp repeat region of PN5; GPII-1 was digested with AatII and XbaI to give the remaining 1.3 kb 3' portion of PN5. A portion of the ligation reaction was transformed into *E. coli* Stable 2 cells. Of the 9.6 kb isolates containing all four fragments, HR-1 was sequenced and found to have the desired 5.4 kb sequence. These isolates grew well and showed no tendency to rearrange. The sequence of this engineered version of PN5 is shown in Figures 5A-E (SEQ ID NO: 5).

EXAMPLE 8

HUMAN PN5

An 856 bp clone (Figure 3A, SEQ ID No.: 3) has been isolated from a human dorsal root ganglia (DRG) cDNA library that is most closely related to rat PN5 with 79% identity for the amino acid sequence. The human PN5 sequence spans the region between IIII1 and interdomain III/IV which includes the fast inactivation gate (i.e., IFM) that is located within interdomain III/IV.

The human DRG cDNA library was constructed from lumbar 4 and 5 DRG total RNA that was randomly primed. First strand cDNA was synthesized with SuperScript II reverse transcriptase (GIBCO BRL) and the second strand synthesis with T4 DNA polymerase. EcoRI adaptors were ligated to the ends of the double stranded cDNAs and the fragments cloned into the ZAP II vector (Stratagene). The library was screened with digoxigenin-labeled rat PN3, rat PN1 and human heart hH1 probes. Positive clones were sequenced and compared to known human and rat sodium channel sequences. Only the aforementioned clone was identified as human PN5 sequence.

Channel	% Similarity	% Identity
Human Brain (HBA)	76	69
Human Heart (hH1)	81	74

	Human Atypical Heart	60	52
	Human Skeletal Muscle	80	71
	Human Neuroendocrine	78	71
	Human PN3	77	70
5	Rat PN1	79	72
	Rat PN3	78	71
	Rat PN4	78	70
	Rat PN5	86	79

10 Figure 3B compares the amino acid sequence of the hPN5 fragment with the rat PN5 amino acid sequence in the appropriate region.

EXAMPLE 9

15 TISSUE DISTRIBUTION BY RT-PCR

Brain, spinal cord, DRG, nodose ganglia, superior cervical ganglia, sciatic nerve, heart and skeletal muscle tissue were isolated from anesthetized, normal adult male Sprague-Dawley rats and were stored at -80°C. RNA was isolated from each tissue using RNAzol (Tel-Test, Inc.). Random-primed cDNA was reverse transcribed from 500ng of RNA from each tissue. The forward primer (CAGATTGTGTTCTCAGTACATTCC) and the reverse primer (CCAGGTGTCTAACGAATAAATAGG) were designed from the 3'-untranslated region to yield a 252 base pair fragment. The cycle parameters were: 94°C/2 min. (denaturation), 94°C/30 sec., 65°C/30 sec. and 72°C/1min. (35 cycles) and 72°C/4 min. The reaction products were analyzed on a 4% agarose gel.

25 A positive control and a no-template control were also included. cDNA from each tissue was also PCR amplified using primers specific for glyceraldehyde-3-phosphate dehydrogenase to demonstrate template viability, as described by Tso *et al.*, Nucleic Acid Res. 13, 2485-2502 (1985).

30 Tissue distribution profile of rPN5 by analysis of RNA from selected rat tissues by RT-PCR was as follows:

<u>Tissue</u>	<u>RT-PCR (35 cycles)</u>
Brain	+

5	Spinal cord	+
	DRG	+++
	Nodose ganglia	+++
	Superior cervical ganglia	+
	Sciatic nerve	-
	Heart	-
	Skeletal muscle	-
	F11-untreated	+
	F11-treated	+

10 PN5 was also detected after only 25 cycles (24 + 1) in the same five tissues as above in the same relative abundance.

EXAMPLE 10

ANTIBODIES

15 A synthetic peptide (26 amino acids in interdomain II and III - residues 977 to 1002) was conjugated to KLH and antibody raised in rabbits. The antiserum was subsequently affinity purified.

PN5 constitutes a subfamily of novel sodium channel genes; these genes are different from those detectable with other probes (e.g., PEA8 and PN3 probes).

20 Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity and understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claims.

SEQUENCE LISTING

(1) 1) GENERAL INFORMATION:

(i) APPLICANT:

- (A) NAME: F. HOFFMANN-LA ROCHE AG
- (B) STREET: Grenzacherstrasse 124
- (C) CITY: Basle
- (D) STATE: BS
- (E) COUNTRY: Switzerland
- (F) POSTAL CODE (ZIP): CH-4010
- (G) TELEPHONE: 061-6884256
- (H) TELEFAX: 061-6881395
- (I) TELEX: 962292/965542 hlr ch

(ii) TITLE OF INVENTION: Nucleic Acid Encoding a Nervous Tissue Sodium Channel

(iii) NUMBER OF SEQUENCES: 5

(iv) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: Floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: PatentIn Release # 1.0, Version # 1.30

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 5908 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: rat
- (F) TISSUE TYPE: Dorsal root ganglia
- (G) CELL TYPE: Peripheral nerve

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

1 GAAGTCACAG GAGTGTCTGT CAGCGAGAGG AAGAAGGGAG AGTTTACTGA
51 GTGTCTTCTG CCCCTCCTCA GGGTGAAGAT GGAGGAGAGG TACTACCCGG

101 TGATCTTCCC GGACGAGCGG AATTTCCGCC CCTTCACTTC CGACTCTCTG
 151 GCTGCCATAG AGAAGCGGAT TGCTATCCAA AAGGAGAGGA AGAAGTCCAA
 201 AGACAAGGCG GCAGCTGAGC CCCAGCCTCG GCCTCAGCTT GACCTAAAGG
 251 CCTCCAGGAA GTTACCTAAG CTTTATGGTG ACATTCCCCC TGAGCTTGTA
 301 GCGAAGCCTC TGGAAGACCT GGACCCATTC TACAAAGACC ATAAGACATT
 351 CATGGTGTG AACAAGAAGA GAACAATTTA TCGCTTCAGC GCCAAGCGGG
 401 CCTTGTTTCAT TCTGGGGCCT TTTAATCCCC TCAGAAGCTT AATGATTTCGT
 451 ATCTCTGTCC ATTCAGTCTT TAGCATGTTC ATCATCTGCA CGGTGATCAT
 501 CAACTGTATG TTCATGGCGA ATTCTATGGA GAGAAGTTTC GACAACGACA
 551 TTCCCGAATA CGTCTTCATT GGGATTTATA TTTTAGAAGC TGTGATTAAA
 601 ATATTGGCAA GAGGCTTCAT TGTGGATGAG TTTTCCTTCC TCCGAGATCC
 651 GTGGAAC TGG GCTTCA TTGTCATTGG AACAGCGATC GCAACTTGTT
 701 TTCCGGGCAG CCAAGTCAAT CTTTCAGCTC TTCGTACCTT CCGAGTGTTT
 751 AGAGCTCTGA AGGCGATTTT AGTTATCTCA GGTCTGAAGG TCATCGTAGG
 801 TGCCCTGCTG CGCTCGGTGA AGAAGCTGGT AGACGTGATG GTCCTCACTC
 851 TCTTCTGCCT CAGCATCTTT GCCCTGGTCG GTCAGCAGCT GTTCATGGGA
 901 ATTCTGAACC AGAAGTGTAT TAAGCACAAC TGTGGCCCCA ACCCTGCATC
 951 CAACAAGGAT TGCTTTGAAA AGGAAAAAGA TAGCGAAGAC TTCATAATGT
 1001 GTGGTACCTG GCTCGGCAGC AGACCCTGTC CCAATGGTTC TACGTGCGAT
 1051 AAAACCACAT TGAACCCAGA CAATAATTAT ACAAAGTTTG ACAACTTTGG
 1101 CTGGTCCTTT CTCGCCATGT TCCGGGTTAT GACTCAAGAC TCCTGGGAGA
 1151 GGCTTTACCG ACAGATCCTG CGGACCTCTG GGATCTACTT TGTCTTCTTC
 1201 TTCGTGGTGG TCATCTTCCT GGGCTCCTTC TACCTGCTTA ACCTAACCCT
 1251 GGCTGTTGTC ACCATGGCTT ATGAAGAACA GAACAGAAAT GTAGCTGCTG
 1301 AGACAGAGGC CAAGGAGAAA ATGTTTCAGG AAGCCCAGCA GCTGTTAAGG
 1351 GAGGAGAAGG AGGCTCTGGT TGCCATGGGA ATTGACAGAA GTTCCCTTAA
 1401 TTCCCTTCAA GCTTCATCCT TTTCCCCGAA GAAGAGGAAG TTTTTCGGTA

1451 GTAAGACAAG AAAGTCCTTC TTTATGAGAG GGTCCAAGAC GGCCCAAGCC
 1501 TCAGCGTCTG ATTCAGAGGA CGATGCCTCT AAAAATCCAC AGCTCCTTGA
 1551 GCAGACCAA CGACTGTCCC AGAACTTGCC AGTGGATCTC TTTGATGAGC
 1601 ACGTGGACCC CCTCCACAGG CAGAGAGCGC TGAGCGCTGT CAGTATCTTA
 1651 ACCATCACCA TGCAGGAACA AGAAAAATTC CAGGAGCCTT GTTTCCTATG
 1701 TGGGAAAAAT TTGGCCTCTA AGTACCTGGT GTGGGACTGT AGCCCTCAGT
 1751 GGCTGTGCAT AAAGAAGGTC CTGCGGACCA TCATGACGGA TCCCTTTACT
 1801 GAGCTGGCCA TCACCATCTG CATCATCATC AATACCGTTT TCTTAGCCGT
 1851 GGAGCACCAC AACATGGATG ACAACTTAAA GACCATACTG AAAATAGGAA
 1901 ACTGGGTTTT CACGGGAATT TTCATAGCGG AAATGTGTCT CAAGATCATC
 1951 GCGCTCGACC CTTACCACTA CTTCCGGCAC GGCTGGAATG TTTTGTACAG
 2001 CATCGTGGCC CTCCTGAGTC TCGCTGATGT GCTCTACAAC AACTGTCTCTG
 2051 ATAACAATAG GTCTTTCTTG GCTTCCCTCA GAGTGCTGAG GGTCTTCAAG
 2101 TTAGCCAAAT CCTGGCCCAC GTTAAACACT CTCATTAAGA TCATCGGCCA
 2151 CTCCGTGGGC GCGCTTGGA ACCTGACTGT GGTCTGACT ATCGTGGTCT
 2201 TCATCTTTTC TGTGGTGGGC ATGCGGCTCT TCGGCACCAA GTTTAACAAG
 2251 ACCGCCTACG CCACCCAGGA GCGGCCCAGG CGGCGCTGGC ACATGGATAA
 2301 TTTCTACCAC TCCTTCCTGG TGGTGTTCG CATCCTCTGT GGGGAATGGA
 2351 TCGAGAACAT GTGGGGCTGC ATGCAGGATA TGGACGGCTC CCCGTTGTGC
 2401 ATCATTGTCT TTGTCCTGAT AATGGTGATC GGGAAGCTTG TGGTGCTTAA
 2451 CCTCTTCATT GCCTTGCTGC TCAATTCCTT CAGCAATGAG GAGAAGGATG
 2501 GGAGCCTGGA AGGAGAGACC AGGAAAACCA AAGTGCAGCT AGCCCTGGAT
 2551 CGGTTCCGCC GGGCCTTCTC CTTGATGCTG CACGCTCTTC AGAGTTTTTG
 2601 TTGCAAGAAA TGCAGGAGGA AAAACTCGCC AAAGCCAAAA GAGACAACAG
 2651 AAAGCTTTGC TGGTGAGAAT AAAGACTCAA TCCTCCCGGA TGCGAGGCCC
 2701 TGGAAGGAGT ATGATACAGA CATGGCTTTG TAACTGGAC AGGCCGGGGC
 2751 TCCGCTGGCC CCACTCGCAG AGGTAGAGGA CGATGTGGAA TATTGTGGTG
 2801 AAGGCGGTGC CCTACCCACC TCACAACATA GTGCTGGAGT TCAGGCCGGT

2851 GACCTCCCTC CAGAGACCAA GCAGCTCACT AGCCCGGATG ACCAAGGGGT
 2901 TGAAATGGAA GTATTTTCTG AAGAAGATCT GCATTTAAGC ATACAGAGTC
 2951 CTCGAAAGAA GTCTGACGCA GTGAGCATGC TCTCGGAATG CAGCACAATT
 3001 GACCTGAATG ATATCTTTAG AAATTTACAG AAAACAGTTT CCCCCAAAAA
 3051 GCAGCCAGAT AGATGCTTTC CCAAGGGCCT TAGTTGTCAC TTTCTATGCC
 3101 ACAAACAGA CAAGAGAAAG TCCCCCTGGG TCCTGTGGTG GAACATTCCG
 3151 AAAACCTGCT ACCAAATCGT GAAGCACAGC TGGTTTGAGA GTTTCATAAT
 3201 CTTTGTTATT CTGCTGAGCA GTGGAGCGCT GATATTTGAA GATGTCAATC
 3251 TCCCCAGCCG GCCCCAAGTT GAGAAATTAC TAAGGTGTAC CGATAATATT
 3301 TTCACATTTA TTTTCCTCCT GGAAATGATC CTGAAGTGGG TGGCCTTTGG
 3351 ATTCCGGAGG TATTTACCA GTGCCTGGTG CTGGCTTGAT TTCCTCATTG
 3401 TGGTGGTGTC TGTGCTCAGT CTCATGAATC TACCAAGCTT GAAGTCCTTC
 3451 CGGACTCTGC GGGCCCTGAG ACCTCTGCGG GCGCTGTCCC AGTTTGAAGG
 3501 AATGAAGGTT GTCGTCTACG CCCTGATCAG CGCCATACCT GCCATTCTCA
 3551 ATGTCTTGCT GGTCTGCCTC ATTTTCTGGC TCGTATTTTG TATCTTGGGA
 3601 GTAAATTTAT TTTCTGGGAA GTTTGGAAGG TGCATTAACG GGACAGACAT
 3651 AAATATGTAT TTGGATTTTA CCGAAGTTCC GAACCGAAGC CAATGTAACA
 3701 TTAGTAATTA CTCGTGGAAG GTCCCGCAGG TCAACTTTGA CAACGTGGGG
 3751 AATGCCTATC TCGCCCTGCT GCAAGTGGCA ACCTATAAGG GCTGGCTGGA
 3801 AATCATGAAT GCTGCTGTCG ATTCCAGAGA GAAAGACGAG CAGCCGGACT
 3851 TTGAGGCGAA CCTCTACGCG TATCTCTACT TTGTGGTTTT TATCATCTTC
 3901 GGCTCCTTCT TTACCCTGAA CCTCTTTATC GGTGTTATTA TTGACAACTT
 3951 CAATCAGCAG CAGAAAAAGT TAGGTGGCCA AGACATTTTT ATGACAGAAG
 4001 AACAGAAGAA ATATTACAAT GCAATGAAAA AGTTAGGAAC CAAGAAACCT
 4051 CAAAAGCCCA TCCCAAGGCC CCTGAACAAA TGTCAAGCCT TTGTGTTCTG
 4101 CCTGGTCACA AGCCAGGTCT TTGACGTCAT CATCTGGGT CTTATTGTCT
 4151 TAAATATGAT TATCATGATG GCTGAATCTG CCGACCAGCC CAAAGATGTG

4201 AAGAAAACCT TTGATATCCT CAACATAGCC TTCGTGGTCA TCTTTACCAT
 4251 AGAGTGTCTC ATCAAAGTCT TTGCTTTGAG GCAACACTAC TTCACCAATG
 4301 GCTGGAACCT ATTTGATTGT GTGGTCGTGG TTCTTTCTAT CATTAGTACC
 4351 CTGGTTTCCC GCTTGGAGGA CAGTGACATT TCTTTCCCGC CCACGCTCTT
 4401 CAGAGTCGTC CGCTTGGCTC GGATTGGTCG AATCCTCAGG CTGGTCCGGG
 4451 CTGCCCCGGG AATCAGGACC CTCCTCTTTG CTTTGATGAT GTCTCTCCCC
 4501 TCTCTCTTCA ACATCGGTCT GCTGCTCTTC CTGGTGATGT TCATTTACGC
 4551 CATCTTTGGG ATGAGCTGGT TTTCCAAAGT GAAGAAGGGC TCCGGGATCG
 4601 ACGACATCTT CAACTTCGAG ACCTTTACGG GCAGCATGCT GTGCCTCTTC
 4651 CAGATAACCA CTTCGGCTGG CTGGGATACC CTCCTCAACC CCATGCTGGA
 4701 GGCAAAGAA CACTGCAACT CCTCCTCCCA AGACAGCTGT CAGCAGCCGC
 4751 AGATAGCCGT CGTCTACTTC GTCAGTTACA TCATCATCTC CTTCTCATC
 4801 GTGGTCAACA TGTACATCGC TGTGATCCTC GAGAACTTCA ACACAGCCAC
 4851 GGAGGAGAGC GAGGACCCTC TGGGAGAGGA CGACTTTGAA ATCTTCTATG
 4901 AGGTCTGGGA GAAGTTTGAC CCCGAGGCGT CGCAGTTCAT CCAGTATTCG
 4951 GCCCTCTCTG ACTTTGCGGA CGCCCTGCCG GAGCCGTTGC GTGTGGCCAA
 5001 GCCGAATAAG TTTCAGTTTC TAGTGATGGA CTTGCCCATG GTGATGGGCG
 5051 ACCGCCTCCA TTGCATGGAT GTTCTCTTTG CTTTCACTAC CAGGGTCCTC
 5101 GGGGACTCCA GCGGCTTGGA TACCATGAAA ACCATGATGG AGGAGAAGTT
 5151 TATGGAGGCC AACCCTTTTA AGAAGCTCTA CGAGCCCATA GTCACCACCA
 5201 CCAAGAGGAA GGAGGAGGAG CAAGGCGCCG CCGTCATCCA GAGGGCCTAC
 5251 CGGAAACACA TGGAGAAGAT GGTCAAACCTG AGGCTGAAGG ACAGGTCAAG
 5301 TTCATCGCAC CAGGTGTTTT GCAATGGAGA CTTGTCCAGC TTGGATGTGG
 5351 CCAAGGTCAA GGTTCAACAAT GACTGAACCC TCATCTCCAC CCCTACCTCA
 5401 CTGCCTCACA GCTTAGCCTC CAGCCTCTGG CGAGCAGGCG GCAGACTCAC
 5451 TGAACACAGG CCGTTCGATC TGTGTTTTTG GCTGAACGAG GTGACAGGTT
 5501 GGCGTCCATT TTAAATGAC TCTTGAAAG ATTCATGTA GAGAGATGTT
 5551 AGAAGGGACT GCAAAGGACA CCGACCATAA CGGAAGGCCT GGAGGACAGT

Gly Pro Phe Asn Pro Leu Arg Ser Leu Met Ile Arg Ile Ser Val His
 115 120 125
 Ser Val Phe Ser Met Phe Ile Ile Cys Thr Val Ile Ile Asn Cys Met
 130 135 140
 Phe Met Ala Asn Ser Met Glu Arg Ser Phe Asp Asn Asp Ile Pro Glu
 145 150 155 160
 Tyr Val Phe Ile Gly Ile Tyr Ile Leu Glu Ala Val Ile Lys Ile Leu
 165 170 175
 Ala Arg Gly Phe Ile Val Asp Glu Phe Ser Phe Leu Arg Asp Pro Trp
 180 185 190
 Asn Trp Leu Asp Phe Ile Val Ile Gly Thr Ala Ile Ala Thr Cys Phe
 195 200 205
 Pro Gly Ser Gln Val Asn Leu Ser Ala Leu Arg Thr Phe Arg Val Phe
 210 215 220
 Arg Ala Leu Lys Ala Ile Ser Val Ile Ser Gly Leu Lys Val Ile Val
 225 230 235 240
 Gly Ala Leu Leu Arg Ser Val Lys Lys Leu Val Asp Val Met Val Leu
 245 250 255
 Thr Leu Phe Cys Leu Ser Ile Phe Ala Leu Val Gly Gln Gln Leu Phe
 260 265 270
 Met Gly Ile Leu Asn Gln Lys Cys Ile Lys His Asn Cys Gly Pro Asn
 275 280 285
 Pro Ala Ser Asn Lys Asp Cys Phe Glu Lys Glu Lys Asp Ser Glu Asp
 290 295 300
 Phe Ile Met Cys Gly Thr Trp Leu Gly Ser Arg Pro Cys Pro Asn Gly
 305 310 315 320
 Ser Thr Cys Asp Lys Thr Thr Leu Asn Pro Asp Asn Asn Tyr Thr Lys
 325 330 335
 Phe Asp Asn Phe Gly Trp Ser Phe Leu Ala Met Phe Arg Val Met Thr
 340 345 350
 Gln Asp Ser Trp Glu Arg Leu Tyr Arg Gln Ile Leu Arg Thr Ser Gly
 355 360 365
 Ile Tyr Phe Val Phe Phe Phe Val Val Val Ile Phe Leu Gly Ser Phe
 370 375 380
 Tyr Leu Leu Asn Leu Thr Leu Ala Val Val Thr Met Ala Tyr Glu Glu
 385 390 395 400
 Gln Asn Arg Asn Val Ala Ala Glu Thr Glu Ala Lys Glu Lys Met Phe

405	410	415
Gln Glu Ala Gln Gln Leu Leu Arg Glu Glu Lys Glu Ala Leu Val Ala		
420	425	430
Met Gly Ile Asp Arg Ser Ser Leu Asn Ser Leu Gln Ala Ser Ser Phe		
435	440	445
Ser Pro Lys Lys Arg Lys Phe Phe Gly Ser Lys Thr Arg Lys Ser Phe		
450	455	460
Phe Met Arg Gly Ser Lys Thr Ala Gln Ala Ser Ala Ser Asp Ser Glu		
465	470	475
Asp Asp Ala Ser Lys Asn Pro Gln Leu Leu Glu Gln Thr Lys Arg Leu		
485	490	495
Ser Gln Asn Leu Pro Val Asp Leu Phe Asp Glu His Val Asp Pro Leu		
500	505	510
His Arg Gln Arg Ala Leu Ser Ala Val Ser Ile Leu Thr Ile Thr Met		
515	520	525
Gln Glu Gln Glu Lys Phe Gln Glu Pro Cys Phe Pro Cys Gly Lys Asn		
530	535	540
Leu Ala Ser Lys Tyr Leu Val Trp Asp Cys Ser Pro Gln Trp Leu Cys		
545	550	555
Ile Lys Lys Val Leu Arg Thr Ile Met Thr Asp Pro Phe Thr Glu Leu		
565	570	575
Ala Ile Thr Ile Cys Ile Ile Ile Asn Thr Val Phe Leu Ala Val Glu		
580	585	590
His His Asn Met Asp Asp Asn Leu Lys Thr Ile Leu Lys Ile Gly Asn		
595	600	605
Trp Val Phe Thr Gly Ile Phe Ile Ala Glu Met Cys Leu Lys Ile Ile		
610	615	620
Ala Leu Asp Pro Tyr His Tyr Phe Arg His Gly Trp Asn Val Phe Asp		
625	630	635
Ser Ile Val Ala Leu Leu Ser Leu Ala Asp Val Leu Tyr Asn Thr Leu		
645	650	655
Ser Asp Asn Asn Arg Ser Phe Leu Ala Ser Leu Arg Val Leu Arg Val		
660	665	670
Phe Lys Leu Ala Lys Ser Trp Pro Thr Leu Asn Thr Leu Ile Lys Ile		
675	680	685
Ile Gly His Ser Val Gly Ala Leu Gly Asn Leu Thr Val Val Leu Thr		
690	695	700

Ile Val Val Phe Ile Phe Ser Val Val Gly Met Arg Leu Phe Gly Thr
 705 710 715 720
 Lys Phe Asn Lys Thr Ala Tyr Ala Thr Gln Glu Arg Pro Arg Arg Arg
 725 730 735
 Trp His Met Asp Asn Phe Tyr His Ser Phe Leu Val Val Phe Arg Ile
 740 745 750
 Leu Cys Gly Glu Trp Ile Glu Asn Met Trp Gly Cys Met Gln Asp Met
 755 760 765
 Asp Gly Ser Pro Leu Cys Ile Ile Val Phe Val Leu Ile Met Val Ile
 770 775 780
 Gly Lys Leu Val Val Leu Asn Leu Phe Ile Ala Leu Leu Leu Asn Ser
 785 790 795 800
 Phe Ser Asn Glu Glu Lys Asp Gly Ser Leu Glu Gly Glu Thr Arg Lys
 805 810 815
 Thr Lys Val Gln Leu Ala Leu Asp Arg Phe Arg Arg Ala Phe Ser Phe
 820 825 830
 Met Leu His Ala Leu Gln Ser Phe Cys Cys Lys Lys Cys Arg Arg Lys
 835 840 845
 Asn Ser Pro Lys Pro Lys Glu Thr Thr Glu Ser Phe Ala Gly Glu Asn
 850 855 860
 Lys Asp Ser Ile Leu Pro Asp Ala Arg Pro Trp Lys Glu Tyr Asp Thr
 865 870 875 880
 Asp Met Ala Leu Tyr Thr Gly Gln Ala Gly Ala Pro Leu Ala Pro Leu
 885 890 895
 Ala Glu Val Glu Asp Asp Val Glu Tyr Cys Gly Glu Gly Gly Ala Leu
 900 905 910
 Pro Thr Ser Gln His Ser Ala Gly Val Gln Ala Gly Asp Leu Pro Pro
 915 920 925
 Glu Thr Lys Gln Leu Thr Ser Pro Asp Asp Gln Gly Val Glu Met Glu
 930 935 940
 Val Phe Ser Glu Glu Asp Leu His Leu Ser Ile Gln Ser Pro Arg Lys
 945 950 955 960
 Lys Ser Asp Ala Val Ser Met Leu Ser Glu Cys Ser Thr Ile Asp Leu
 965 970 975
 Asn Asp Ile Phe Arg Asn Leu Gln Lys Thr Val Ser Pro Lys Lys Gln
 980 985 990
 Pro Asp Arg Cys Phe Pro Lys Gly Leu Ser Cys His Phe Leu Cys His

995	1000	1005
Lys Thr Asp Lys Arg Lys Ser Pro Trp Val Leu Trp Trp Asn Ile Arg		
1010	1015	1020
Lys Thr Cys Tyr Gln Ile Val Lys His Ser Trp Phe Glu Ser Phe Ile		
1025	1030	1035 1040
Ile Phe Val Ile Leu Leu Ser Ser Gly Ala Leu Ile Phe Glu Asp Val		
1045	1050	1055
Asn Leu Pro Ser Arg Pro Gln Val Glu Lys Leu Leu Arg Cys Thr Asp		
1060	1065	1070
Asn Ile Phe Thr Phe Ile Phe Leu Leu Glu Met Ile Leu Lys Trp Val		
1075	1080	1085
Ala Phe Gly Phe Arg Arg Tyr Phe Thr Ser Ala Trp Cys Trp Leu Asp		
1090	1095	1100
Phe Leu Ile Val Val Val Ser Val Leu Ser Leu Met Asn Leu Pro Ser		
1105	1110	1115 1120
Leu Lys Ser Phe Arg Thr Leu Arg Ala Leu Arg Pro Leu Arg Ala Leu		
1125	1130	1135
Ser Gln Phe Glu Gly Met Lys Val Val Val Tyr Ala Leu Ile Ser Ala		
1140	1145	1150
Ile Pro Ala Ile Leu Asn Val Leu Leu Val Cys Leu Ile Phe Trp Leu		
1155	1160	1165
Val Phe Cys Ile Leu Gly Val Asn Leu Phe Ser Gly Lys Phe Gly Arg		
1170	1175	1180
Cys Ile Asn Gly Thr Asp Ile Asn Met Tyr Leu Asp Phe Thr Glu Val		
1185	1190	1195 1200
Pro Asn Arg Ser Gln Cys Asn Ile Ser Asn Tyr Ser Trp Lys Val Pro		
1205	1210	1215
Gln Val Asn Phe Asp Asn Val Gly Asn Ala Tyr Leu Ala Leu Leu Gln		
1220	1225	1230
Val Ala Thr Tyr Lys Gly Trp Leu Glu Ile Met Asn Ala Ala Val Asp		
1235	1240	1245
Ser Arg Glu Lys Asp Glu Gln Pro Asp Phe Glu Ala Asn Leu Tyr Ala		
1250	1255	1260
Tyr Leu Tyr Phe Val Val Phe Ile Ile Phe Gly Ser Phe Phe Thr Leu		
1265	1270	1275 1280
Asn Leu Phe Ile Gly Val Ile Ile Asp Asn Phe Asn Gln Gln Lys		
1285	1290	1295

Lys Leu Gly Gly Gln Asp Ile Phe Met Thr Glu Glu Gln Lys Lys Tyr
 1300 1305 1310
 Tyr Asn Ala Met Lys Lys Leu Gly Thr Lys Lys Pro Gln Lys Pro Ile
 1315 1320 1325
 Pro Arg Pro Leu Asn Lys Cys Gln Ala Phe Val Phe Asp Leu Val Thr
 1330 1335 1340
 Ser Gln Val Phe Asp Val Ile Ile Leu Gly Leu Ile Val Leu Asn Met
 1345 1350 1355 1360
 Ile Ile Met Met Ala Glu Ser Ala Asp Gln Pro Lys Asp Val Lys Lys
 1365 1370 1375
 Thr Phe Asp Ile Leu Asn Ile Ala Phe Val Val Ile Phe Thr Ile Glu
 1380 1385 1390
 Cys Leu Ile Lys Val Phe Ala Leu Arg Gln His Tyr Phe Thr Asn Gly
 1395 1400 1405
 Trp Asn Leu Phe Asp Cys Val Val Val Val Leu Ser Ile Ile Ser Thr
 1410 1415 1420
 Leu Val Ser Arg Leu Glu Asp Ser Asp Ile Ser Phe Pro Pro Thr Leu
 1425 1430 1435 1440
 Phe Arg Val Val Arg Leu Ala Arg Ile Gly Arg Ile Leu Arg Leu Val
 1445 1450 1455
 Arg Ala Ala Arg Gly Ile Arg Thr Leu Leu Phe Ala Leu Met Met Ser
 1460 1465 1470
 Leu Pro Ser Leu Phe Asn Ile Gly Leu Leu Leu Phe Leu Val Met Phe
 1475 1480 1485
 Ile Tyr Ala Ile Phe Gly Met Ser Trp Phe Ser Lys Val Lys Lys Gly
 1490 1495 1500
 Ser Gly Ile Asp Asp Ile Phe Asn Phe Glu Thr Phe Thr Gly Ser Met
 1505 1510 1515 1520
 Leu Cys Leu Phe Gln Ile Thr Thr Ser Ala Gly Trp Asp Thr Leu Leu
 1525 1530 1535
 Asn Pro Met Leu Glu Ala Lys Glu His Cys Asn Ser Ser Ser Gln Asp
 1540 1545 1550
 Ser Cys Gln Gln Pro Gln Ile Ala Val Val Tyr Phe Val Ser Tyr Ile
 1555 1560 1565
 Ile Ile Ser Phe Leu Ile Val Val Asn Met Tyr Ile Ala Val Ile Leu
 1570 1575 1580
 Glu Asn Phe Asn Thr Ala Thr Glu Glu Ser Glu Asp Pro Leu Gly Glu

1585	1590	1595	1600
Asp Asp Phe Glu Ile Phe Tyr Glu Val Trp Glu Lys Phe Asp Pro Glu			
1605	1610	1615	
Ala Ser Gln Phe Ile Gln Tyr Ser Ala Leu Ser Asp Phe Ala Asp Ala			
1620	1625	1630	
Leu Pro Glu Pro Leu Arg Val Ala Lys Pro Asn Lys Phe Gln Phe Leu			
1635	1640	1645	
Val Met Asp Leu Pro Met Val Met Gly Asp Arg Leu His Cys Met Asp			
1650	1655	1660	
Val Leu Phe Ala Phe Thr Thr Arg Val Leu Gly Asp Ser Ser Gly Leu			
1665	1670	1675	1680
Asp Thr Met Lys Thr Met Met Glu Glu Lys Phe Met Glu Ala Asn Pro			
1685	1690	1695	
Phe Lys Lys Leu Tyr Glu Pro Ile Val Thr Thr Thr Lys Arg Lys Glu			
1700	1705	1710	
Glu Glu Gln Gly Ala Ala Val Ile Gln Arg Ala Tyr Arg Lys His Met			
1715	1720	1725	
Glu Lys Met Val Lys Leu Arg Leu Lys Asp Arg Ser Ser Ser Ser His			
1730	1735	1740	
Gln Val Phe Cys Asn Gly Asp Leu Ser Ser Leu Asp Val Ala Lys Val			
1745	1750	1755	1760
Lys Val His Asn Asp			
1765			

(4) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 856 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: human
- (F) TISSUE TYPE: Dorsal root ganglia
- (G) CELL TYPE: Peripheral nerve

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

1 GCTGAGCAGT GGGGCACTGA TATTTGAAGA TGTTACCTT GAGAACCAAC

51 CCAAATCCA AGAATTACTA AATTGTACTG ACATTATTTT TACACATATT
 101 TTTATCCTGG AGATGGTACT AAAATGGGTA GCCTTCGGAT TTGGAAAGTA
 151 TTTCACCAGT GCCTGGTGCT GCCTTGATTT CATCATTGTG ATTGTCTCTG
 201 TGACCACCCT CATTAACCTA ATGGAATTGA AGTCCTTCCG GACTCTACGA
 251 GCACTGAGGC CTCTTCGTGC GCTGTCCCAG TTTGAAGGAA TGAAGGTGGT
 301 GGTCAATGCT CTCATAGGTG CCATACCTGC CATTCTGAAT GTTTTGCTTG
 351 TCTGCCTCAT TTTCTGGCTC GTATTTTGTA TTCTGGGAGT ATACTTCTTT
 401 TCTGGAAAAT TTGGGAAATG CATTAATGGA ACAGACTCAG TTATAAATTA
 451 TACCATCATT ACAAATAAAA GTCAATGTGA AAGTGGCAAT TTCTCTTGGA
 501 TCAACCAGAA AGTCAACTTT GACAATGTGG GAAATGCTTA CCTCGCTCTG
 551 CTGCAAGTGG CAACATTTAA GGGCTGGATG GATATTATAT ATGCAGCTGT
 601 TGATTCCACA GAGAAAGAAC AACAGCCAGA GTTTGAGAGC AATTCACCTG
 651 GTTACATTTA CTTCGTAGTC TTTATCATCT TTGGCTCATT CTTCACTCTG
 701 AATCTCTTCA TTGGCGTTAT CATTGACAAC TTCAACCAAC AGCAGAAAAA
 751 GTTAGGTGGC CAAGACATTT TTATGACAGA AGAACAGAAG AAATACTATA
 801 ATGCAATGAA AAAATTAGGA TCCAAAAAAC CTCAAAAACC CATTCCACGG
 851 CCCGTT

(5) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 702 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: RT-PCR

- (A) DESCRIPTION: /desc = „DNA probe/domain IV“

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: rat
- (F) TISSUE TYPE: dorsal root ganglia
- (G) CELL TYPE: peripheral nerve

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

```
1   CTCAACATGG TTACGATGAT GGTGGAGACC GACGAGCAGG GCGAGGAGAA
51  GACGAAGGTT CTGGGCAGAA TCAACCAGTT CTTTGTGGCC GTCTTCACGG
101 GCGAGTGTGT GATGAAGATG TTCGCCCTGC GACAGTACTA TTTCACCAAC
151 GGCTGGAACG TGTTCGACTT CATAGTGGTG ATCCTGTCCA TTGGGAGTCT
201 GCTGTTTCT  GCAATCCTTA AGTCACTGGA AACTACTTC  TCCCCGACGC
251 TCTTCCGGGT CATCCGTCTG GCCAGGATCG GCCGCATCCT CAGGCTGATC
301 CGAGCAGCCA AGGGGATTCG CACGCTGCTC TTCGCCCTCA TGATGTCCCT
351 GCCCCCCTC TTCAACATCG GCCTCCTCCT CTCCTCGTC  ATGTTCATCT
401 ACTCCATCTT CGGCATGGCC AGCTTCGCTA ACGTCGTGGA CGAGGCCGGC
451 ATCGACGACA TGTTCAACTT CAAGACCTTT GGCAACAGCA TGCTGTGCCT
501 GTTCCAGATC ACCACCTCGG CCGGCTGGGA CGGCCTCCTC AGCCCCATCC
551 TCAACACGGG GCCTCCCTAC TGCACCCCA ACCTGCCCAA CAGCAACGGC
601 TCCCGGGGGA ACTGCGGGAG CCCGGCGGTG GGCATCATCT TCTTACCAC
651 CTACATCATC ATCTCCTTCC TCATCGTGGT CAACATGTAT ATCGCAGTCA
701 TC
```

(5) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 5334 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single

- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: RT-PCR
- (A) DESCRIPTION: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
- (A) ORGANISM:
- (F) TISSUE TYPE:
- (G) CELL TYPE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

```

1  GTCGACTCTA GATCAGGGTG AAGATGGAGG AGAGGTACTA CCCGGTGATC
51  TTCCCGGACG AGCGGAATTT CCGCCCCTTC ACTTCCGACT CTCTGGCTGC
101 CATAGAGAAG CGGATTGCTA TCCAAAAGGA GAGGAAGAAG TCCAAAGACA
151 AGGCGGCAGC TGAGCCCCAG CCTCGGCCTC AGCTTGACCT AAAGGCCTCC
201 AGGAAGTTAC CTAAGCTTTA TGGTGACATT CCCCCTGAGC TTGTAGCGAA
251 GCCTCTGGAA GACCTGGACC CATTCTACAA AGACCATAAG ACATTCATGG
301 TGTGTAACAA GAAGAGAACA ATTTATCGCT TCAGCGCCAA GCGGGCCTTG
351 TTCATTCTGG GGCCTTTTAA TCCCCTCAGA AGCTTAATGA TTCGTATCTC
401 TGTCCATTCA GTCTTTAGCA TGTTCATCAT CTGCACGGTG ATCATCAACT
451 GTATGTTTCA GCGGAATTCT ATGGAGAGAA GTTTCGACAA CGACATTCCC
501 GAATACGTCT TCATTGGGAT TTATATTTTA GAAGCTGTGA TTAAAATATT
551 GGCAAGAGGC TTCATTGTGG ATGAGTTTTT CTCCTCCGA GATCCGTGGA
601 ACTGGCTGGA CTTCATTTGTC ATTGGAACAG CGATCGCAAC TTGTTTTCCG
651 GGCAGCCAAG TCAATCTTTC AGCTCTTCGT ACCTTCCGAG TG TTCAGAGC
701 TCTGAAGGCG ATTTTCAGTTA TCTCAGGTCT GAAGGTCATC GTAGGTGCCC
751 TGCTGCGCTC GGTGAAGAAG CTGGTAGACG TGATGGTCCT CACTCTCTTC
801 TGCCTCAGCA TCTTTGCCCT GGTGCGTCAG CAGCTGTTCA TGGGAATTCT
851 GAACCAGAAG TGTATTAAGC ACAACTGTGG CCCCAACCCT GCATCCAACA
901 AGGATTGCTT TGAAAAGGAA AAAGATAGCG AAGACTTCAT AATGTGTGGT
951 ACCTGGCTCG GCAGCAGACC CTGTCCCAAT GGTCTACGT GCGATAAAAC

```

1001 CACATTGAAC CCAGACAATA ATTATACAAA GTTTGACAAC TTTGGCTGGT
 1051 CCTTTCTCGC CATGTTCCGG GTTATGACTC AAGACTCCTG GGAGAGGCTT
 1101 TACCGACAGA TCCTGCGGAC CTCTGGGATC TACTTTGTCT TCTTCTTCGT
 1151 GGTGGTCATC TTCCTGGGCT CCTTCTACCT GCTTAACCTA ACCCTGGCTG
 1201 TTGTCACCAT GGCTTATGAA GAACAGAACA GAAATGTAGC TGCTGAGACA
 1251 GAGGCCAAGG AGAAAATGTT TCAGGAAGCC CAGCAGCTGT TAAGGGAGGA
 1301 GAAGGAGGCT CTGGTTGCCA TGGGAATTGA CAGAAGTTCC CTTAATTCCC
 1351 TTCAAGCTTC ATCCTTTTCC CCGAAGAAGA GGAAGTTTTT CGGTAGTAAG
 1401 ACAAGAAAGT CCTTCTTTAT GAGAGGGTCC AAGACGGCCC AAGCCTCAGC
 1451 GTCTGATTCA GAGGACGATG CCTCTAAAAA TCCACAGCTC CTTGAGCAGA
 1501 CCAAACGACT GTCCCAGAAC TTGCCAGTGG ATCTCTTTGA TGAGCACGTG
 1551 GACCCCCTCC ACAGGCAGAG AGCGCTGAGC GCTGTCAGTA TCTTAACCAT
 1601 CACCATGCAG GAACAAGAAA AATTCCAGGA GCCTTGTTTC CCATGTGGGA
 1651 AAAATTTGGC CTCTAAGTAC CTGGTGTGGG ACTGTAGCCC TCAGTGGCTG
 1701 TGCATAAAGA AGGTCCTGCG GACCATCATG ACGGATCCCT TTACTGAGCT
 1751 GGCCATCACC ATCTGCATCA TCATCAATAC CGTTTTCTTA GCCGTGGAGC
 1801 ACCACAACAT GGATGACAAC TTAAAGACCA TACTGAAAAT AGGAAACTGG
 1851 GTTTTCACGG GAATTTTCAT AGCGGAAATG TGTCTCAAGA TCATCGCGCT
 1901 CGACCCTTAC CACTACTTCC GGCACGGCTG GAATGTTTTT GACAGCATCG
 1951 TGGCCCTCCT GAGTCTCGCT GATGTGCTCT ACAACACACT GTCTGATAAC
 2001 AATAGGTCTT TCTTGGCTTC CCTCAGAGTG CTGAGGGTCT TCAAGTTAGC
 2051 CAAATCCTGG CCCACGTAA ACACTCTCAT TAAGATCATC GGCCACTCCG
 2101 TGGGCGCGCT TGGAAACCTG ACTGTGGTCC TGACTATCGT GGTCTTCATC
 2151 TTTTCTGTGG TGGGCATGCG GCTCTTCGGC ACCAAGTTTA ACAAGACCGC
 2201 CTACGCCACC CAGGAGCGGC CCAGGCGGCG CTGGCACATG GATAATTTCT
 2251 ACCACTCCTT CCTGGTGGTG TTCCGCATCC TCTGTGGGGA ATGGATCGAG
 2301 AACATGTGGG GCTGCATGCA GGATATGGAC GGCTCCCCGT TGTGCATCAT
 2351 TGTCTTTGTC CTGATAATGG TGATCGGGAA GCTTGTGGTG CTTAACCTCT

2401 TCATTGCCTT GCTGCTCAAT TCCTTCAGCA ATGAGGAGAA GGATGGGAGC
 2451 CTGGAAGGAG AGACCAGGAA AACCAAAGTG CAGCTAGCCC TGGATCGGTT
 2501 CCGCCGGGCC TTCTCCTTCA TGCTGCACGC TCTTCAGAGT TTTTGTGCA
 2551 AGAAATGCAG GAGGAAAAAC TCGCCAAAGC CAAAAGAGAC AACAGAAAGC
 2601 TTTGCTGGTG AGAATAAAGA CTCAATCCTC CCGGATGCGA GGCCCTGGAA
 2651 GGAGTATGAT ACAGACATGG CTTTGTACAC TGGACAGGCC GGGGCTCCGC
 2701 TGGCCCCACT CGCAGAGGTA GAGGACGATG TGAATATTG TGGTGAAGGC
 2751 GGTGCCCTAC CCACCTCACA ACATAGTGCT GGAGTTCAGG CCGGTGACCT
 2801 CCCTCCAGAG ACCAAGCAGC TCACTAGCCC GGATGACCAA GGGGTTGAAA
 2851 TGGAAGTATT TTCTGAAGAA GATCTGCATT TAAGCATACA GAGTCCTCGA
 2901 AAGAAGTCTG ACGCAGTGAG CATGCTCTCG GAATGCAGCA CAATTGACCT
 2951 GAATGATATC TTTAGAAATT TACAGAAAAC AGTTTCCCCC AAAAAGCAGC
 3001 CAGATAGATG CTTTCCCAAG GGCCTTAGTT GTCACTTTCT ATGCCACAAA
 3051 ACAGACAAGA GAAAGTCCCC CTGGGTCTCTG TGGTGGAACA TTCGAAAAAC
 3101 CTGCTACCAA ATCGTGAAGC ACAGCTGGTT TGAGAGTTTC ATAATCTTTG
 3151 TTATTCTGCT GAGCAGTGGA GCGCTGATAT TTGAAGATGT CAATCTCCCC
 3201 AGCCGGCCCC AAGTTGAGAA ATTACTAAGG TGTACCGATA ATATTTTCAC
 3251 ATTTATTTTC CTCCTGGAAC TGATCCTGAA GTGGGTGGCC TTTGGATTCC
 3301 GGAGGTATTT CACCAGTGCC TGGTGCTGGC TTGATTTCTT CATTGTGGTG
 2251 GTGTCTGTGC TCAGTCTCAT GAATCTACCA AGCTTGAAGT CCTTCCGGAC
 3401 TCTGCGGGCC CTGAGACCTC TGCGGGCGCT GTCCCAGTTT GAAGGAATGA
 3451 AGGTTGTCGT CTACGCCCTG ATCAGCGCCA TACCTGCCAT TCTCAATGTC
 3501 TTGCTGGTCT GCCTCATTTT CTGGCTCGTA TTTTGTATCT TGGGAGTAAA
 3551 TTTATTTTCT GGGAAGTTTG GAAGGTGCAT TAACGGGACA GACATAAATA
 3601 TGTATTTTGA TTTTACCGAA GTTCCGAACC GAAGCCAATG TAACATTAGT
 3651 AATTACTCGT GGAAGGTCCC GCAGGTCAAC TTTGACAACG TGGGGAATGC
 3701 CTATCTCGCC CTGCTGCAAG TGGCAACCTA TAAGGGCTGG CTGGAAATCA
 3751 TGAATGCTGC TGTCGATTCC AGAGAGAAAG ACGAGCAGCC GGACTTTGAG

3801 GCGAACCTCT ACGCGTATCT CTACTTTGTG GTTTTTATCA TCTTCGGCTC
 3851 CTTCTTTACC CTGAACCTCT TTATCGGTGT TATTATTGAC AACTTCAATC
 3901 AGCAGCAGAA AAAGTTAGGT GGCCAAGACA TCTTCATGAC TGAGGAGCAG
 3951 AAGAAATATT ACAATGCAAT GAAAAAGTTA GGAACCAAGA AACCTCAAAA
 4001 GCCCATCCCA AGGCCCTGA ACAAATGTCA AGCCTTTGTG TTCGACCTGG
 4051 TCACAAGCCA GGTCTTTGAC GTCATCATTC TGGGTCTTAT TGTCTTAAAT
 4101 ATGATTATCA TGATGGCTGA ATCTGCCGAC CAGCCCAAAG ATGTGAAGAA
 4151 AACCTTTGAT ATCCTCAACA TAGCCTTCGT GGTCACTTTT ACCATAGAGT
 4201 GTCTCATCAA AGTCTTTGCT TTGAGGCAAC ACTACTTCAC CAATGGCTGG
 4251 AACTTATTTG ATTGTGTGGT CGTGGTTCTT TCTATCATTG GTACCCTGGT
 4301 TTCCCGCTTG GAGGACAGTG ACATTTCTTT CCCGCCACG CTCTTCAGAG
 4351 TCGTCCGCTT GGCTCGGATT GGTCAATCC TCAGGCTGGT CCGGGCTGCC
 4401 CGGGGAATCA GGACCCTCCT CTTTGCTTTG ATGATGTCTC TCCC TCTCT
 4451 CTTCAACATC GGTCTGCTGC TCTTCCTGGT GATGTTCAAT TACGCCATCT
 4501 TTGGGATGAG CTGGTTTTCC AAAGTGAAGA AGGGCTCCGG GATCGACGAC
 4551 ATCTTCAACT TCGAGACCTT TACGGGCAGC ATGCTGTGCC TCTTCCAGAT
 4601 AACCACCTCG GCTGGCTGGG ATACCCTCCT CAACCCCATG CTGGAGGCAA
 4651 AAGAACACTG CAACTCCTCC TCCAAGACA GCTGTCAGCA GCCGCAGATA
 4701 GCCGTCGTCT ACTTCGTCAG TTACATCATC ATCTCCTTCC TCATCGTGGT
 4751 CAACATGTAC ATCGCTGTGA TCCTCGAGAA CTTCAACACA GCCACGGAGG
 4801 AGAGCGAGGA CCCTCTGGGA GAGGACGACT TTGAAATCTT CTATGAGGTC
 4851 TGGGAGAAGT TTGACCCCGA GCGTCGCAG TTCATCCAGT ATTCCGGCCCT
 4901 CTCTGACTTT GCGGACGCCC TGCCGGAGCC GTTGCCTGTG GCCAAGCCGA
 4951 ATAAGTTTCA GTTTCTAGTG ATGGACTTGC CCATGGTGAT GGGCGACCGC
 5001 CTCCATTGCA TGGATGTTCT CTTTGCTTTC ACTACCAGGG TCCTCGGGGA
 5051 CTCCAGCGGC TTGGATACCA TGAAAACCAT GATGGAGGAG AAGTTTATGG
 5101 AGGCCAACCC TTTTAAGAAG CTCTACGAGC CCATAGTCAC CACCACCAAG
 5151 AGGAAGGAGG AGGAGCAAGG CGCCGCCGTC ATCCAGAGGG CCTACCGGAA

5201 ACACATGGAG AAGATGGTCA AACTGAGGCT GAAGGACAGG TCAAGTTCAT
5251 CGCACCAGGT GTTTTGCAAT GGAGACTTGT CCAGCTTGGA TGTGGCCAAG
5301 GTCAAGGTTT ACAATGACTG AACCCTCATC TAGA

CLAIMS

What is claimed is:

1. An isolated DNA sequence comprising the nucleotide sequence set forth in SEQ ID NO:1 or SEQ ID NO:3.
2. The DNA of Claim 1 wherein said DNA sequence is encoding a sodium channel protein or fragment thereof.
3. The DNA of Claim 2 wherein said sodium channel protein is the α -subunit or fragment thereof.
4. The DNA of Claim 3 wherein said sodium channel protein is tetrodotoxin-resistant.
5. The DNA of Claim 3 or 4 wherein said sodium channel protein is found in mammals.
6. The DNA of Claim 3 or 4 wherein said sodium channel protein is found in rat.
7. The DNA of Claim 3 or 4 wherein said sodium channel protein is found in human.
8. The DNA of Claim 1 wherein said DNA is cDNA.
9. The DNA of Claim 1 wherein said DNA is synthetic DNA.
10. Expression vectors comprising the DNA of Claim 8.
11. Expression vectors comprising the synthetic DNA of Claim 9.
12. Host cells transformed with the expression vectors of Claim 10.
13. Host cells transformed with the expression vectors of Claim 11.
14. A recombinant polynucleotide comprising a nucleic acid sequence derived from the DNA sequence of Claim 1.
15. A sodium channel protein encoded by a DNA of Claims 1 to 9 or allelic variants thereof.
16. A tetrodotoxin-resistant sodium channel protein encoded by a DNA of Claims 1 to 9 or allelic variants thereof.
17. The protein of Claim 16 having the amino acid sequence set forth in SEQ ID NO:2.
18. A method for identifying inhibitors of tetrodotoxin-resistant sodium channel protein comprising contacting a compound suspected of being said inhibitor with sodium channel protein of claim 16 and measuring the activity of said expressed sodium channel protein.
19. Poly- and/or monoclonal antibodies raised against a tetrodotoxin-resistant sodium channel protein encoded by a DNA of Claims 1 to 9 or allelic variants thereof.
20. A diagnostic kit comprising a polynucleotide of claim 14 capable of specifically hybridizing to a tetrodotoxin-resistant sodium channel protein or fragment thereof.
21. The use of an isolated DNA sequence of Claims 1 to 9 for identifying a compound suspected of being an inhibitor of tetrodotoxin-resistant sodium channel protein.
22. The invention substantially as hereinbefore described especially with reference to the foregoing Examples.



Application No: GB 9825378.4
Claims searched: 1-22

Examiner: Dr J Houlihan
Date of search: 29 April 1999

Patents Act 1977
Search Report under Section 17

Databases searched:

UK Patent Office collections, including GB, EP, WO & US patent specifications, in:
UK Cl (Ed.Q):
Int Cl (Ed.6):
Other: ONLINE: WPI, EPODOC, PAJ, CAS ONLINE, DGENE, BIOSCIENCE/STN

Documents considered to be relevant:

Category	Identity of document and relevant passage	Relevant to claims
X	WO 97/01577 A1 (UNI. COLL. LONDON) page 2 lines 10-20; Examples 1 & 2	1 at least
X	WO 96/14077 A1 (TROPHIX PHARM. INC.) Whole document	1 at least
X	Gene Vol. 202 1997. Chen J <i>et.al.</i> "Molecular cloning of a putative tetrodotoxin-resistant sodium channel from dog nodose ganglion neurons" pages 7-14	1 at least

X	Document indicating lack of novelty or inventive step	A	Document indicating technological background and/or state of the art.
Y	Document indicating lack of inventive step if combined with one or more other documents of same category.	P	Document published on or after the declared priority date but before the filing date of this invention.
&	Member of the same patent family	E	Patent document published on or after, but with priority date earlier than, the filing date of this application.